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(54) Title: CLONING AND EXPRESSION OF cDNA FOR HUMAN DIHYDROPYRIMIDINE DEHYDROGENASE (57) Abstract <p>The invention relates to methods and compositions that are useful for detecting deficiencies in dihydropyrimidine dehydrogenase (DPD) levels in mammals including humans. Cancer patients having a DPD deficiency are at risk of a severe toxic reaction to the commonly used anticancer agent 5-fluorouracil (5-FU). Claimed are DPD genes from human and pig, methods for detecting the level of nucleic acids that encode DPD in a patient, and nucleic acids that are useful as probes for this purpose. Also claimed are methods for expressing DPD in heterologous organisms. Expression vectors that employ a DPD nucleic acid as a selectable marker are also claimed. This selectable marker functions in both prokaryotes and eukaryotes.</p>		

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5 CLONING AND EXPRESSION OF cDNA FOR HUMAN
 DIHYDROPYRIMIDINE DEHYDROGENASE

 TECHNICAL FIELD OF THE INVENTION

 The present invention relates to methods and compositions for
detecting deficiencies in dihydropyrimidine dehydrogenase (DPD) levels in
10 mammals, including humans. The methods and compositions are useful for
identifying persons who are at risk of a toxic reaction to the commonly
employed cancer chemotherapy agent 5-fluorouracil.

 BACKGROUND OF THE INVENTION

15 5-Fluorouracil (5-FU) is commonly used in the treatment of
cancers, including cancers of the breast, head, neck, and digestive system.
The efficacy of 5-FU as a cancer treatment varies significantly among patients.
Clinically significant differences in systemic clearance and systemic exposure of
5-FU are often observed. [Grem, J.L. *In* Chabner, B.A. and J.M. Collins (eds.),
20 *Cancer Chemotherapy: Principles and Practice*, pp. 180-224, Philadelphia, PA,
Lippincott, 1990)]. Furthermore, 5-FU treatment is severely toxic to some
patients, and has even caused death. [Fleming *et al.* (1993) *Eur. J. Cancer*
29A: 740-744; Thyss *et al.* (1986) *Cancer Chemother. Pharmacol.* 16: 64-66;
Santini *et al.* (1989) *Br. J. Cancer* 59: 287-290; Goldberg *et al.* (1988) *Br. J.*
25 *Cancer* 57: 186-189; Trump *et al.* (1991) *J. Clin. Oncol.* 9: 2027-2035; Au *et*
al. (1982) *Cancer Res.* 42: 2930-2937].

 Patients in whom 5-FU is severely toxic typically have low levels
of dihydropyrimidine dehydrogenase (DPD) activity [Tuchman *et al.* (1985) *N.*
Engl. J. Med. 313: 245-249; Diasio *et al.* (1988) *J. Clin. Invest.* 81: 47-51;
30 Fleming *et al.* (1991) *Proc. Am. Assoc. Cancer Res.* 32: 179; Harris *et al.*
(1991) *Cancer (Phila.)* 68: 499-501; Houyau *et al.* (1993) *J. Nat'l. Cancer Inst.*
85: 1602-1603; Lyss *et al.* (1993) *Cancer Invest.* 11: 239-240].
Dihydropyrimidine dehydrogenase (DPD, EC 1.3.1.2) is the principal enzyme
involved in the degradation of 5-FU, which acts by inhibiting thymidylate
35 synthase [Heggie *et al.* (1987) *Cancer Res.* 47: 2203-2206; Chabner *et al.*

(1989) In DeVita *et al.* (eds.), *Cancer - Principles and Practice of Oncology*, pp. 349-395, Philadelphia, PA, Lippincott; Diasio *et al.* (1989) *Clin. Pharmacokinet* 16: 215-237; Grem *et al.*, *supra.*). The level of DPD activity also affects the efficacy of 5-FU treatments, as 5-FU plasma levels are inversely correlated with the level of DPD activity [Iigo *et al.* (1988) *Biochem. Pharm.* 37: 1609-1613; Goldberg *et al.*, *supra.*; Harris *et al.*, *supra.*; Fleming *et al.*, *supra.*]. In turn, the efficacy of 5-FU treatment of cancer is correlated with plasma levels of 5-FU.

In addition to its 5-FU degrading activity, DPD is also the initial and rate limiting enzyme in the three-step pathway of uracil and thymine catabolism, leading to the formation of β -alanine and β -aminobutyric acid, respectively [Wasternack *et al.* (1980) *Pharm. Ther.* 8: 629-665]. DPD deficiency is associated with inherited disorders of pyrimidine metabolism, clinically termed thymine-uraciluria [Bakkeren *et al.* (1984) *Clin. Chim. Acta.* 140: 247-256]. Clinical symptoms of DPD deficiency include a nonspecific cerebral dysfunction, and DPD deficiency is associated with psychomotor retardation, convulsions, and epileptic conditions [Berger *et al.* (1984) *Clin. Chim. Acta* 141: 227-234; Wadman *et al.* (1985) *Adv. Exp. Med. Biol.* 165A: 109-114; Wilcken *et al.* (1985) *J. Inherit. Metab. Dis.* 8 (Suppl. 2): 115-116; van Gennip *et al.* (1989) *Adv. Exp. Med. Biol.* 253A: 111-118; Brockstedt *et al.* (1990) *J. Inherit. Metab. Dis.* 12: 121-124; Duran *et al.* (1991) *J. Inherit. Metab. Dis.* 14: 367-370]. Biochemically, patients having DPD deficiency have an almost complete absence of DPD activity in fibroblasts [Bakkeren *et al.*, *supra.*] and in lymphocytes [Berger *et al.*, *supra.*; Piper *et al.* (1980) *Biochim. Biophys. Acta* 633: 400-409]. These patients typically have a large accumulation of uracil and thymine in their cerebrospinal fluid [Bakkeren *et al.*, *supra.*] and urine [Berger *et al.*, *supra.*; Bakkeren *et al.*, *supra.*; Brockstedt *et al.*, *supra.*; Fleming *et al.* (1992) *Cancer Res.* 52: 2899-2902].

Familial studies suggest that DPD deficiency follows an autosomal recessive pattern of inheritance [Diasio *et al.*, (1988) *supra.*]. Up to three percent of the general human population are estimated to be putative heterozygotes for DPD deficiency, as determined by enzymatic activity in lymphocytes [Milano and Eteinne (1994) *Pharmacogenetics* (in press)]. This suggests that the frequency of homozygotes for DPD deficiency may be as high as one person per thousand.

DPD has been purified from liver tissue of rats [Shiotani and Weber (1981) *J. Biol. Chem.* 256: 219-224; Fujimoto *et al.* (1991); *J. Nutr. Sci. Vitaminol.*

37: 89-98], pig [Podschun *et al.* (1989) *Eur. J. Biochem.* 185: 219-224], cattle [Porter *et al.* (1991) *J. Biol. Chem.* 266: 19988-19994], and human [Lu *et al.* (1992) *J. Biol. Chem.* 267: 1702-1709]. The pig enzyme contains flavins and iron-sulfur prosthetic groups and exists as a homodimer with a monomer Mr of about 107,000 [Podschun *et al.*, *supra.*]. Since the enzyme exhibits a nonclassical two-site ping-pong mechanism, it appears to have distinct binding sites for NADPH/NADP and uracil/5,6-dihydrouracil [Podschun *et al.* (1990) *J. Biol. Chem.* 265: 12966-12972]. An acid-base catalytic mechanism has been proposed for DPD [Podschun *et al.* (1993) *J. Biol. Chem.* 268: 3407-3413].

Because an undetected DPD deficiency poses a significant danger to a cancer patient who is being treated with 5-FU, a great need exists for a simple and accurate test for DPD deficiency. Such a test will also facilitate diagnosis of disorders that are associated with DPD deficiency, such as uraciluria. The present invention provides such a test, thus fulfilling these and other needs.

SUMMARY OF THE INVENTION

The claimed invention includes isolated nucleic acids that code for a dihydropyrimidine dehydrogenase (DPD) protein. Human and pig DPD cDNA sequences are claimed (Seq. ID No. 1 and Seq. ID No. 3, respectively), as are DPD nucleic acids that are capable of selectively hybridizing to the human or pig DPD cDNAs under stringent hybridization conditions. Oligonucleotide probes that are capable of selectively hybridizing, under stringent hybridizing conditions, to a human or pig DPD nucleic acid are also claimed. The invention also includes isolated nucleic acids that code for a DPD polypeptide that specifically binds to an antibody generated against an immunogen consisting of a human or pig DPD polypeptide having an amino acid sequence as depicted by Seq. ID No. 2 or Seq. ID No. 4.

Also claimed are methods for determining whether a patient is at risk of a toxic reaction to 5-fluorouracil (5-FU). The methods involve analyzing DPD DNA or mRNA in a sample from the patient to determine the amount of intact DPD nucleic acid. An enhanced risk of a toxic reaction to 5-fluorouracil is indicated by a decrease in the amount of intact DPD DNA or mRNA in the sample compared to the amount of DPD DNA or mRNA in a sample obtained from a patient known to not

have a DPD deficiency, or by a defect in the DPD nucleic acid that results in an inadequate level of DPD activity.

The invention also includes methods for expressing recombinant DPD protein in a prokaryotic cell. The methods involve transfecting the cell with an expression vector comprising a promoter that is operably linked to a nucleic acid that encodes DPD, and incubating the cell in a medium that contains uracil to allow expression of the recombinant DPD protein.

Also claimed are expression vectors that utilize a nucleic acid that encodes DPD as a selectable marker. These selectable markers function in both eukaryotes and prokaryotes.

BRIEF DESCRIPTION OF THE FIGURES

Figures 1A-1B show the nucleotide sequence of the human *DPYD* cDNA.

Figures 2A-2B shows the nucleotide sequence of the pig *DPYD* cDNA.

Figure 3 shows a comparison of the pig and human DPD cDNA deduced amino acid sequences. Only those amino acid residues of human DPD that differ from the pig sequences are shown *below* the pig DPD amino acid sequence. The following motifs relevant for catalytic activity are *boxed*: NADPH/NADP binding, FAD binding, uracil binding, and 4Fe-4S binding.

Figure 4 shows the pedigree of a family used for a study of inheritance of DPD deficiency. Symbols are as follows: □ male, ○ female. Dotted symbols indicate intermediate DPD activity, a dashed square indicates high (normal) DPD activity, and ■ indicates undetectable DPD activity.

Figure 5 shows a Southern blot of the products from reverse transcriptase PCR amplified cDNA for the subjects shown in Figure 4. The 906 and 741 bp bands correspond to the wild-type and the deleted DPD cDNA fragments, respectively. "+" signifies the presence of the wild-type allele and "-" signifies the presence of the mutant allele.

Figure 6 is a schematic of the wild-type and mutant DPD cDNAs. Numbers above the cDNA graphical representation represent nucleotide positions. Start and stop codons are indicated.

Figure 7 is a PCR analysis of the DPD cDNA deletion found in the subject family. The numbers of the subjects correspond to those indicated in Figure

4. Lane 6 is a negative control (no template present) and Lane 7 contains a 1 kb marker ladder (GIBCO BRL).

5

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Definitions

Abbreviations for the twenty naturally occurring amino acids follow conventional usage. In the polypeptide notation used herein, the left-hand direction is the amino terminal direction and the right-hand direction is the carboxy-terminal direction, in accordance with standard usage and convention.

10

The term "nucleic acids," as used herein, refers to either DNA or RNA. Included are single or double-stranded polymers of deoxyribonucleotide or ribonucleotide bases. Self-replicating plasmids, infectious polymers of DNA or RNA and nonfunctional DNA or RNA are included. Unless specified otherwise, the left hand end of single-stranded polynucleotide sequences is the 5' end. The direction of 5' to 3' addition of ribonucleotides to nascent RNA transcripts is referred to as the transcription direction; sequence regions on the DNA strand having the same sequence as the RNA and which are 5' to the 5' end of the RNA transcript are referred to as "upstream sequences;" sequence regions on the DNA strand having the same sequence as the RNA and which are 3' to the 3' end of the RNA transcript are referred to as "downstream sequences."

15

20

"Nucleic acid probes" or "oligonucleotide probes" can be DNA or RNA fragments. Where a specific sequence for a nucleic acid probe is given, it is understood that the complementary strand is also identified and included. The complementary strand will work equally well in situations where the target is a double-stranded nucleic acid.

25

The phrase "selectively hybridizing to" refers to a nucleic acid probe that, under appropriate hybridization conditions, hybridizes, duplexes or binds only to a particular target DNA or RNA sequence when the target sequences are present in a preparation of DNA or RNA. "Complementary" or "target" nucleic acid sequences refer to those nucleic acids that selectively hybridize to a nucleic acid probe. Proper annealing conditions depend, for example, upon a probe's length, base composition, and the number of mismatches and their position on the probe, and must often be determined empirically. For discussions of nucleic acid probe

30

design and annealing conditions, see, for example, Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (2nd ed.), Vols. 1-3, Cold Spring Harbor Laboratory, (1989) or *Current Protocols in Molecular Biology*, F. Ausubel *et al.*, (ed.) Greene Publishing and Wiley-Interscience, New York (1987).

5 The terms "stringent conditions" and "conditions of high stringency" refer to conditions under which a nucleic acid probe will hybridize substantially to its target subsequence, but to no other sequences. Stringent conditions are sequence-dependent and will be different in different circumstances. Longer sequences hybridize specifically at higher temperatures. Generally, stringent
10 conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a complementary probe. Typically, stringent conditions will be those in which the salt concentration is at least about 0.2 molar at pH 7 and the
15 temperature is at least about 60°C for long sequences (e.g. greater than about 50 nucleotides) and at least about 42°C for shorter sequences (e.g. 10 to 50 nucleotides). As other factors may significantly affect the stringency of hybridization, including, among others, base composition and size of the complementary strands, the presence of organic solvents and the extent of base
20 mismatching, the combination of parameters is more important than the absolute measure of any one.

A nucleic acid is said to "encode" or "code for" a specific protein when the nucleic acid sequence comprises, in the proper order, codons for each of the amino acids of the protein or a specific subsequence of the protein. The nucleic
25 acids include both the DNA strand that is transcribed into RNA and the RNA strand that is translated into protein. It is further understood that the invention includes nucleic acids that differ from the DPD sequences specifically disclosed herein in that particular codons are replaced by degenerate codons, so that the variant nucleic acid encodes a protein having the same amino acid sequence as that
30 encoded by the specifically disclosed nucleic acids.

The phrase "isolated" or "substantially pure," when referring to nucleic acids that encode DPD, refers to nucleic acids that are sufficiently pure that the predominant nucleic acid species in the preparation is the desired DPD nucleic

acid. Preferably, the DPD nucleic acids are more than 70% pure, more preferably greater than 90% pure, and most preferably greater than 95% pure.

The term "control sequence" refers to a DNA sequence or sequences that are capable, when properly attached to a desired coding sequence, of causing expression of the coding sequence. Such control sequences include at least promoters and, optionally, transcription termination signals. Additional factors necessary or helpful for expression can also be included. As used herein, "control sequences" simply refers to whatever DNA sequence signal that is useful to result in expression in the particular host used. Often, control sequences are utilized as an "expression cassette," in which the control sequences are operably linked to the nucleic acid that is to be expressed.

The term "operably linked" as used herein refers to a juxtaposition wherein the components are configured so as to perform their usual function. Thus, control sequences or promoters operably linked to a coding sequence are capable of effecting the expression of the coding sequence.

The term "vector" refers to nucleic acids that are capable of replicating in a selected host organism. The vector can replicate as an autonomous structure, or alternatively can integrate into the host cell chromosome(s) and thus replicate along with the host cell genome. Vectors include viral- or bacteriophage-based expression systems, autonomous self-replicating circular DNA (plasmids), and include both expression and nonexpression vectors. The term "plasmid" refers to an autonomous circular DNA molecule capable of replication in a cell, and includes both the expression and nonexpression types.

The phrase "recombinant protein" or "recombinantly produced protein" refers to a peptide or protein produced using recombinant DNA techniques. Host cells produce the recombinant protein because they have been genetically altered by the introduction of the appropriate nucleic acid that codes for the protein. Typically, the heterologous nucleic acid is introduced as part of an expression vector.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: "reference sequence", "comparison window", "sequence identity", "percentage of sequence identity", and "substantial identity". A "reference sequence" is a defined sequence used as a basis for a sequence comparison; a reference sequence can comprise a complete

cDNA or gene sequence, such as the nucleic acid sequence of Seq. ID Nos. 1 or 3, or can be a subset of a larger sequence, for example, as a segment of a full-length cDNA or gene sequence.

Optimal alignment of sequences for aligning a comparison window can
5 be conducted by the local homology algorithm of Smith and Waterman (1981) *Adv. Appl. Math.* 2:482, by the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. (USA)* 85:2444, or by computerized
10 implementations of these algorithms (*e.g.*, GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Dr., Madison, WI).

The terms "substantial identity" or "substantial sequence identity" as applied to nucleic acids and as used herein denote a characteristic of a nucleotide sequence wherein the polynucleotide comprises a sequence that has at least 85
15 percent sequence identity, preferably at least 90 to 95 percent sequence identity, and more preferably at least 99 percent sequence identity as compared to a reference sequence over a comparison window of at least 20 nucleotide positions, frequently over a window of at least 25-50 nucleotides. The percentage of sequence identity is calculated by comparing the reference sequence to the
20 polynucleotide sequence, which may include deletions or additions which total 20 percent or less of the reference sequence over the window of comparison. The reference sequence may be a subset of a larger sequence, such as a segment or subsequence of the human DPD gene disclosed herein.

As applied to polypeptides, the terms "substantial identity" or
25 "substantial sequence identity" mean that two peptide sequences, when optimally aligned, such as by the programs GAP or BESTFIT using default gap weights, share at least 80 percent sequence identity, preferably at least 90 percent sequence identity, more preferably at least 95 percent sequence identity or more. "Percentage amino acid identity" or "percentage amino acid sequence identity"
30 refers to a comparison of the amino acids of two polypeptides which, when optimally aligned, have approximately the designated percentage of the same amino acids. For example, "95% amino acid identity" refers to a comparison of the amino acids of two polypeptides which when optimally aligned have 95% amino acid identity. Preferably, residue positions that are not identical differ by conservative

amino acid substitutions. For example, the substitution of amino acids having similar chemical properties such as charge or polarity are not likely to effect the properties of a protein. Examples include glutamine for asparagine or glutamic acid for aspartic acid.

5 The phrase "substantially purified" or "isolated" when referring to a DPD polypeptide means a chemical composition that is essentially free of other cellular components. The DPD polypeptide is preferably in a homogeneous state, although it can be in either a dry form or in an aqueous solution. Purity and homogeneity are typically determined using analytical chemistry techniques such as
10 polyacrylamide gel electrophoresis (PAGE) or high performance liquid chromatography (HPLC). A protein that is the predominant species present in a preparation is considered substantially purified. Generally, a substantially purified or isolated protein will comprise more than 80% of all macromolecular species present in the preparation. Preferably, the protein is purified to represent greater than 90%
15 of all macromolecular species present. More preferably the protein is purified to greater than 95%, and most preferably the protein is purified to essential homogeneity, wherein other macromolecular species are not detected by conventional techniques.

20 The phrase "specifically binds to an antibody" or "specifically immunoreactive with," when referring to a protein or peptide, refers to a binding reaction that is determinative of the presence of the protein in the presence of a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies bind to a particular protein and do not bind in a significant amount to other proteins present in the sample. Obtaining
25 an antibody that specifically binds to a particular protein may require screening. For example, antibodies raised to the human DPD protein immunogen with the amino acid sequence depicted in SEQ. ID No. 2 can be selected to obtain antibodies specifically immunoreactive with DPD proteins and not with other proteins. These antibodies recognize proteins that are homologous to the human DPD protein, such
30 as DPD proteins from other mammalian species. A variety of immunoassay formats can be used to select antibodies specifically immunoreactive with a particular protein. For example, solid-phase enzyme-linked immunoassays (ELISAs) are routinely used to select monoclonal antibodies specifically immunoreactive with a protein. See Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring

Harbor Publications, New York, for a description of immunoassay formats and conditions that can be used to determine specific immunoreactivity.

5 Detailed Description of the Preferred Embodiment

10 The claimed invention provides compositions and methods that are useful for detecting deficient or diminished DPD activity in mammals, including humans. These methods and compositions are useful for identifying people who are at risk of a toxic reaction to the chemotherapy agent 5-fluorouracil. Methods and compositions for treating mammals who suffer from an insufficient level of DPD are also provided. Also included in the invention are methods for expressing high levels of DPD in prokaryotes, and selectable markers that function in both prokaryotes and eukaryotes.

15 The claimed methods and compositions are based on the discovery of an isolated cDNA that codes for human dihydropyrimidine dehydrogenase (DPD). A newly discovered cDNA that codes for pig DPD is also described. The human (SEQ. ID No. 1) and pig (SEQ. ID No. 3) DPD cDNA sequences are presented in Figures 1A-1B and 2A-2B, respectively. An alignment of the human and pig DPD deduced amino acid sequences is shown in Figure 3. The nucleic acids of the invention are useful for determining whether a patient has an abnormal DPD gene, or whether the DPD gene in a patient is expressed an insufficient level. Either of these conditions can result in a DPD deficiency that can cause the patient to be susceptible to 5-FU toxicity. By detecting the DPD deficiency before treatment commences, the clinician can either adjust the dose of 5-FU downward, or can choose an alternative chemotherapy agent.

25 A. Description and Isolation of DPD Nucleic Acids

1. Description of DPD Nucleic Acids

30 The nucleic acids of the invention are typically identical to or show substantial sequence identity (determined as described above) to the nucleic acid sequences of SEQ ID No. 1 or SEQ ID No. 3. Nucleic acids encoding human DPD will typically hybridize to the nucleic acid sequence of SEQ ID Nos. 1 or 3 under stringent hybridization conditions as described herein.

Also claimed are isolated nucleic acids that code for a DPD polypeptide that specifically binds to an antibody generated against a specific immunogen, such as an immunogen that has of the amino acid sequence depicted by SEQ ID Nos. 2 or 4, or a specific subsequence of these polypeptides. To
5 identify whether a nucleic acid encodes such a DPD polypeptide, an immunoassay is typically employed. Typically, the immunoassay will use a polyclonal or monoclonal antibody that was raised against the protein of SEQ ID Nos. 2 or 4. The antibody is selected to have low cross-reactivity against other (non-DPD) polypeptides, and any such cross- reactivity is removed by immunoadsorption prior
10 to use in the immunoassay.

In order to produce antisera for use in an immunoassay, the DPD protein of SEQ ID Nos. 2 or 4 is isolated as described herein, for example, by recombinant expression. An inbred strain of mouse such as Balb/c is immunized with the DPD protein using a standard adjuvant, such as Freund's adjuvant, and a
15 standard mouse immunization protocol (see Harlow and Lane, *supra*). Alternatively, a synthetic peptide derived from the amino acid sequences disclosed herein and conjugated to a carrier protein can be used an immunogen. Polyclonal sera are collected and titered against the immunogen protein in an immunoassay, for example, a solid phase immunoassay with the immunogen immobilized on a solid
20 support. Polyclonal antisera with a titer of 10^4 or greater are selected and tested for their cross reactivity against non-DPD proteins, using a competitive binding immunoassay such as the one described in Harlow and Lane, *supra*, at pages 570-573. Three non-DPD proteins are used in this determination: the IRK protein [Kubo
25 *et al.* (1993) *Nature* 362:127], the G-IRK protein [Kubo *et al.* (1993) *Nature* 364:802] and the ROM-K protein [Ho *et al.* (1993) *Nature* 362:127]. These non-DPD proteins can be produced as recombinant proteins and isolated using standard molecular biology and protein chemistry techniques as described herein.

Immunoassays in the competitive binding format can be used for the crossreactivity determinations. For example, the DPD protein of SEQ ID Nos. 2 or 4
30 can be immobilized to a solid support. Proteins added to the assay compete with the binding of the antisera to the immobilized antigen. The ability of the above proteins to compete with the binding of the antisera against the immobilized protein is compared to the DPD protein of Seq. ID Nos. 2 or 4. The percent crossreactivity for the above proteins is calculated, using standard calculations. Those antisera

with less than 10% crossreactivity with each of the proteins listed above are selected and pooled. The cross-reacting antibodies are then removed from the pooled antisera by immunoadsorption with the above-listed proteins.

5 The immunoadsorbed and pooled antisera are then used in a competitive binding immunoassay as described above to determine whether a nucleic acid codes for a DPD polypeptide that specifically binds to an antibody generated against human or pig DPD polypeptide of SEQ ID No. 2 or 4, respectively. The second protein (the protein encoded by the nucleic acid of interest) and the immunogen protein (the human or pig DPD protein of SEQ ID Nos. 10 2 or 4) are compared for their ability to inhibit binding of the antiserum to immobilized human or pig DPD polypeptide. In order to make this comparison, the two proteins are each assayed at a wide range of concentrations to determine the amount of each protein required to inhibit the binding of the antisera to the immobilized protein by 50%. If the amount of the second protein required is less 15 than 10 times the amount of the human DPD protein of SEQ ID No. 2 that is required, then the second protein is said to specifically bind to an antibody generated to an immunogen consisting of the human DPD protein of SEQ ID No. 2. Similarly, the second protein is said to specifically bind to an antibody generated against an immunogen consisting of the pig DPD protein of SEQ ID No. 4 if the 20 amount of second protein required to block antiserum binding by 50% is ten times or less than the amount of pig DPD protein required.

2. Isolation of DPD Nucleic Acids

25 The DPD nucleic acid compositions of this invention, whether cDNA, genomic DNA, RNA, or a hybrid of the various combinations, may be isolated from natural sources or may be synthesized *in vitro*. The nucleic acids claimed can be present in transformed or transfected whole cells, in a transformed or transfected cell lysate, or in a partially purified or substantially pure form.

30 Techniques for manipulating the DPD and other nucleic acids, such as those techniques used for subcloning the nucleic acids into expression vectors, labelling probes, nucleic acid hybridization, and the like are described generally in Sambrook *et al.*, *Molecular Cloning - A Laboratory Manual* (2nd Ed.), Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1989, which is

incorporated herein by reference. This manual is hereinafter referred to as "Sambrook."

Various methods for isolating the DPD nucleic acids are available. For example, one can isolate DNA from a genomic or cDNA library by using labelled
5 oligonucleotide probes that have nucleotide sequences that are complementary to the human and pig DPD gene sequences disclosed herein (SEQ. ID Nos. 1 and 3, respectively). One can use full-length probes or oligonucleotide probes that are based on specific subsequences of these genes. Probes are discussed more fully below. One can use such probes directly in hybridization assays to identify nucleic
10 acids that code for DPD, or one can use amplification methods such as PCR to isolate DPD nucleic acids.

Methods for making and screening cDNA libraries are well known. See, e.g., Gubler, U. and Hoffman, B.J. (1983) *Gene* 25: 263-269 and Sambrook, *supra*. Briefly, to prepare a cDNA library for the purpose of isolating a
15 DPD cDNA, one isolates mRNA from tissue that expresses DPD. Liver is a particularly useful tissue for this purpose, as are peripheral blood lymphocytes. Most other cells also likely produce DPD due to its critical role in pyrimidine degradation and β -alanine synthesis. cDNA is then prepared from the mRNA using standard techniques and ligated into a recombinant vector. The vector is
20 transfected into a recombinant host for propagation, screening and cloning.

Methods for preparing genomic libraries are also well known to those of skill in the art. See, e.g., Sambrook, *supra*. Typically, one can prepare a genomic library by extracting DNA from tissue and either mechanically shearing or enzymatically digesting the DNA to yield fragments of about 12-20kb, or longer if a
25 cosmid is used as the cloning vector. Fragments of the desired size are purified by density gradient centrifugation or gel electrophoresis. The fragments are then cloned into suitable cloning vectors, such as bacteriophage lambda vectors or cosmids. If phage or cosmids are used, one then packages the DNA *in vitro*, as described in Sambrook, *supra*. Recombinant phage or cosmids are analyzed by
30 plaque hybridization as described in Benton and Davis, (1977) *Science* 196: 180-182. Colony hybridization is carried out as generally described in Grunstein *et al.* (1975) *Proc. Natl. Acad. Sci. USA*. 72: 3961-3965.

Standard techniques are used to screen the cDNA or genomic DNA libraries to identify those vectors that contain a nucleic acid that encodes a human

or mammalian DPD. For example, Southern blots are utilized to identify those library members that hybridize to nucleic acid probes derived from the human or pig DPD nucleotide sequences shown in Figures 1A-1B and 2A-2B, respectively. *See, e.g., Sambrook, supra.*

5 Alternatively, one can prepare DPD nucleic acids by using any of various methods of amplifying target sequences, such as the polymerase chain reaction. For example, one can use polymerase chain reaction (PCR) to amplify DPD nucleic acid sequences directly from mRNA, from cDNA or genomic DNA, or from genomic DNA libraries or cDNA libraries. Briefly, to use PCR to isolate the
10 DPD nucleic acids from genomic DNA, one synthesizes oligonucleotide primer pairs that are complementary to the 3' sequences that flank the DNA region to be amplified. One can select primers to amplify the entire region that codes for a full-length DPD polypeptide, or to amplify smaller DNA segments that code for part of the DPD polypeptide, as desired. Suitable primer pairs for amplification of the
15 human *DPYD* gene are shown in Table 1 and are listed as SEQ ID Nos. 5 and 6, 7 and 8, 9 and 10. Polymerase chain reaction is then carried out using the two primers. *See, e.g., PCR Protocols: A Guide to Methods and Applications.* (Innis, M., Gelfand, D., Sninsky, J. and White, T., eds.), Academic Press, San Diego (1990). Amplified fragments can be used as hybridization probes to identify other DPD
20 nucleic acids, such as those from organisms other than human and pig.

Other methods known to those of skill in the art can also be used to isolate DNA encoding the DPD polypeptides. *See, e.g., Sambrook, supra.,* for a description of other techniques that are useful for isolating DNA that codes for specific polypeptides.

25

B. Diagnostic Methods: Detection of DPD Deficiency by Nucleic Acid Detection

To permit the clinician to determine whether a patient has diminished
30 or deficient DPD activity, and thus an enhanced risk of a toxic reaction to 5-FU, the present invention provides methods and reagents for detecting DNA and RNA molecules that code for DPD. These methods permit one to detect DPD deficiency in a patient whether the deficiency is due to a deleted DPD gene (*DPYD*), a DPD gene that is expressed at a lower than normal rate, or a missense or nonsense
35 mutation that results in an abnormal DPD polypeptide. If any of these tests indicate

that the patient has a DPD deficiency, the clinician should exercise extreme caution in using 5-FU as a chemotherapy agent. These methods are also suitable for diagnosing other disorders that are caused by DPD nucleic acid deficiency, such as thymine uraciluria.

5

1. Oligonucleotide Probes

One aspect of the invention is nucleic acid probes that are useful for detecting the presence or absence of DPD nucleic acids in a sample from a human or other mammal. Typically, oligonucleotides are used, although longer fragments that comprise most or all of a DPD gene are also suitable. The claimed probes are specific for human or pig DPD genes. Oligonucleotide probes are generally between about 10 and 100 nucleotides in length, and are capable of selectively hybridizing, under stringent hybridizing conditions, to a target region, a specific subsequence of a DPD nucleic acid. The probes selectively hybridize to DPD nucleic acids, meaning that under stringent hybridization conditions the probes do not substantially hybridize to non-DPD nucleic acids (less than 50% of the probe molecules hybridize to non-DPD nucleic acids). One of skill will recognize that oligonucleotide probes complementary to specific subsequences of the target regions, but not to the entire target region, will also function in the claimed assays so long as such probes selectively hybridize to the target regions.

Alternatively, the oligonucleotide probe can comprise a concatemer that has the formula $[X-Y-Z]_n$, wherein:

a) X is a sequence of 0 to 100 nucleotides or nucleotide analogs that are not complementary to a DPD nucleic acid;

b) Y is a sequence of 10 to 100 nucleotides or nucleotide analogs that are capable of hybridizing under stringent hybridizing conditions to a DPD nucleic acid;

c) Z is a sequence of nucleotides the same as or different from X, such that nucleotides or nucleotide analogs are not complementary to a DPD nucleic acid; and

d) n is 1-500, or more and, where n is greater than 1, Y can be the same or different sequences of nucleotides having the indicated hybridization capability. The probe can be free or contained within a vector sequence (e.g., plasmids or single stranded DNA).

The degree of complementarity (homology) required for detectable binding with the DPD nucleic acids will vary in accordance with the stringency of the hybridization medium and/or wash medium. The degree of complementarity will optimally be 100 percent; however, it should be understood that minor variations in the DPD nucleic acids may be compensated for by reducing the stringency of the hybridization and/or wash medium as described below. Thus, despite the lack of 100 percent complementarity under reduced conditions of stringency, functional probes having minor base differences from their DPD nucleic acid targets are possible. Therefore, under hybridization conditions of reduced stringency, it may be possible to modify up to 60% of a given oligonucleotide probe while maintaining an acceptable degree of specificity. In addition, analogs of nucleosides may be substituted within the probe for naturally occurring nucleosides. This invention is intended to embrace these species when referring to polynucleic acid probes.

Suitable oligonucleotide probes include synthetic oligonucleotides, cloned DNA fragments, PCR products, and RNA molecules. The nature of the probe is not important, provided that it hybridizes specifically to DPD nucleic acids, and not to other nucleic acids under stringent hybridization conditions.

To obtain large quantities of DNA or RNA probes, one can either clone the desired sequence using traditional cloning methods, such as described in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor, New York, 1989, or one can produce the probes by chemical synthesis using commercially available DNA synthesizers. An example of cloning would involve insertion of all or part of the cDNA for the human or pig DPD gene into a replication vector, such as pBR322, M13, or into a vector containing the SP6 promotor (*e.g.*, for generation of single-stranded DPD RNA using SP6 RNA polymerase), and transformation of a bacterial host. The probes can be purified from the host cell by lysis and nucleic acid extraction, treatment with selected restriction enzymes, and further isolation by gel electrophoresis.

Oligonucleotide probes can be chemically synthesized using commercially available methods and equipment. For example, the solid phase phosphoramidite triester method first described by Beaucage and Carruthers [(1981) *Tetrahedron Lett.* 22: 1859-1862] is suitable. This method can be used to produce relatively short probes of between 10 and 50 bases. The triester method described by Matteucci *et al.* [(1981) *J. Am. Chem. Soc.*, 103:3185] is also suitable for

synthesizing oligonucleotide probes. Conveniently, one can use an automated oligonucleotide synthesizer such as the Model 394 DNA/RNA Synthesizer from Applied Biosystems (Foster City, CA) using reagents supplied by the same company.

5 After synthesis, the oligonucleotides are purified either by native acrylamide gel electrophoresis or by anion-exchange HPLC as described in, for example, Pearson and Regnier (1983) *J. Chrom.* 255: 137-149. The sequence of the synthetic oligonucleotide can be verified using the chemical degradation method of Maxam, A.M. and Gilbert, W. (1980) *In* Grossman, L. and Moldave, D., eds.
10 Academic Press, New York, *Methods in Enzymology*, 65:499-560.

 Probes can be comprised of the natural nucleotides or known analogs of the natural nucleotides, including those modified to bind labeling moieties. Oligonucleotide probes that comprise thionucleotides, and thus are resistant to nuclease cleavage, are also suitable. One can use probes that are the full length of
15 the DPD coding regions, or probes that hybridize to a specific subsequence of a DPD gene. Shorter probes are empirically tested for specificity. Preferably, nucleic acid probes are 15 nucleotides or longer in length, although oligonucleotide probe lengths of between about 10 and 100 nucleotides or longer are appropriate. Sambrook, *supra*, describes methods for selecting nucleic acid probe sequences for
20 use in nucleic acid hybridization.

 For purposes of this invention, the probes are typically labelled so that one can detect whether the probe has bound to a DPD nucleic acid. Probes can be labeled by any one of several methods typically used to detect the presence of hybrid polynucleotides. The most common method of detection is the use of
25 autoradiography using probes labeled with ^3H , ^{125}I , ^{35}S , ^{14}C , ^{32}P , or the like. The choice of radioactive isotope depends on research preferences due to ease of synthesis, stability, and half lives of the selected isotopes. Other labels include ligands which bind to antibodies labeled with fluorophores, chemiluminescent agents, and enzymes. Alternatively, probes can be conjugated directly with labels
30 such as fluorophores, chemiluminescent agents or enzymes. The choice of label depends on sensitivity required, ease of conjugation with the probe, stability requirements, and available instrumentation.

 The choice of label dictates the manner in which the label is bound to the probe. Radioactive probes are typically made using commercially available

nucleotides containing the desired radioactive isotope. The radioactive nucleotides can be incorporated into probes, for example, by using DNA synthesizers, by nick translation or primer extension with DNA polymerase I, by tailing radioactive nucleotides to the 3' end of probes with terminal deoxynucleotidyl transferase, by
5 incubating single-stranded M13 plasmids having specific inserts with the Klenow fragment of DNA polymerase in the presence of radioactive deoxynucleotides, dNTP, by transcribing from RNA templates using reverse transcriptase in the presence of radioactive deoxynucleotides, dNTP, or by transcribing RNA from vectors containing specific RNA viral promoters (e.g., SP6 promoter) using the
10 corresponding RNA polymerase (e.g., SP6 RNA polymerase) in the presence of radioactive ribonucleotides rNTP.

The probes can be labeled using radioactive nucleotides in which the isotope resides as a part of the nucleotide molecule, or in which the radioactive component is attached to the nucleotide via a terminal hydroxyl group that has
15 been esterified to a radioactive component such as inorganic acids, e.g., ^{32}P phosphate or ^{14}C organic acids, or esterified to provide a linking group to the label. Base analogs having nucleophilic linking groups, such as primary amino groups, can also be linked to a label.

Non-radioactive probes are often labeled by indirect means. For
20 example, a ligand molecule is covalently bound to the probe. The ligand then binds to an anti-ligand molecule which is either inherently detectable or covalently bound to a detectable signal system, such as an enzyme, a fluorophore, or a chemiluminescent compound. Ligands and anti-ligands may be varied widely. Where a ligand has a natural anti-ligand, namely ligands such as biotin, thyroxine, and cortisol, it
25 can be used in conjunction with its labeled, naturally occurring anti-ligands. Alternatively, any haptenic or antigenic compound can be used in combination with an antibody.

Probes can also be labeled by direct conjugation with a label. For example, cloned DNA probes have been coupled directly to horseradish peroxidase or alkaline phosphatase, as described in Renz, M., and Kurz, K. (1984) A Colorimetric Method for DNA Hybridization. *Nucl. Acids Res.* 12: 3435-3444. Synthetic oligonucleotides have been coupled directly to alkaline phosphatase [Jablonski, E.,
30 *et al.* (1986) Preparation of Oligodeoxynucleotide-Alkaline Phosphatase Conjugates and Their Use as Hybridization Probes. *Nucl. Acids Res.* 14: 6115-6128; and Li P.,

et al. (1987) Enzyme-linked Synthetic Oligonucleotide probes: Non-Radioactive Detection of Enterotoxigenic *Escherichia coli* in Faeca Specimens. *Nucl. Acids Res.* 15: 5275-5287].

Enzymes of interest as labels will typically be hydrolases, such as phosphatases, esterases and glycosidases, or oxidoreductases, particularly peroxidases. Fluorescent compounds include fluorescein and its derivatives, rhodamine and its derivatives, dansyl, umbelliferone, etc. Chemiluminescers include luciferin, and 2,3-dihydrophthalazinediones, *e.g.*, luminol.

The oligonucleotide or polynucleotide acid probes of this invention can be included in a kit which can be used to rapidly determine the level of DPD DNA or mRNA in cells of a human or other mammalian sample. The kit includes all components necessary to assay for the presence of the DPD DNA or mRNA. In the universal concept, the kit includes a stable preparation of labeled probes specific for DPD nucleic acids, hybridization solution in either dry or liquid form for the hybridization of target and probe polynucleotides, as well as a solution for washing and removing undesirable and nonduplexed polynucleotides, a substrate for detecting the labeled duplex, and optionally an instrument for the detection of the label.

The probe components described herein include combinations of probes in dry form, such as lyophilized nucleic acid or in precipitated form, such as alcohol precipitated nucleic acid or in buffered solutions. The label can be any of the labels described above. For example, the probe can be biotinylated using conventional means and the presence of a biotinylated probe can be detected by adding avidin conjugated to an enzyme, such as horseradish peroxidase, which can then be contacted with a substrate which, when reacted with peroxidase, can be monitored visually or by instrumentation using a colorimeter or spectrophotometer. This labeling method and other enzyme-type labels have the advantage of being economical, highly sensitive, and relatively safe compared to radioactive labeling methods. The various reagents for the detection of labeled probes and other miscellaneous materials for the kit, such as instructions, positive and negative controls, and containers for conducting, mixing, and reacting the various components, would complete the assay kit.

2. Assays for Detecting DPD Nucleic Acid Deficiency

One embodiment of the invention provides assays for determining whether a patient is at risk of a toxic reaction to 5-fluorouracil, or suffers from a condition that is caused by inadequate levels of DPD (such as thymine uraciluria). The assay methods involve determining whether the patient is deficient in DPD
5 nucleic acids. A deficiency can arise if the patient is lacking all or part of one or both copies of the DPD gene, or if the DPD gene is not expressed in the appropriate cells of the patient. Another potential cause of DPD deficiency that is detectable using the claimed invention is a nonsense or missense mutation in the DPD gene that results in an abnormal DPD polypeptide.

10 Assay test protocols for use in this invention are those of convention in the field of nucleic acid hybridization, and include both single phase, where the target and probe polynucleic acids are both in solution, and mixed phase hybridizations, where either the target or probe polynucleotides are fixed to an immobile support. The assay test protocols are varied and are not to be considered
15 a limitation of this invention. A general review of hybridization can be had from a reading of *Nucleic Acid Hybridization: A Practical Approach*, Hames and Higgins, eds., IRL Press, 1985; and *Hybridization of Nucleic Acids Immobilized on Solid Supports*, Meinkoth and Wah (1984) *Analytical Biochemistry*, pp. 238, 267-284. Mixed phase hybridizations are preferred.

20 One potential cause of DPD deficiency is a deletion of all or part of one or more copies of the DPD gene in a patient's chromosomal DNA. To determine whether a patient lacks a gene that codes for DPD, the clinician can employ a Southern blot or other means suitable for detecting the presence of a specific nucleotide sequence in genomic DNA. A variety of methods for specific
25 DNA and RNA measurement using nucleic acid hybridization techniques are known to those of skill in the art. See, e.g., Sambrook, *supra*. Briefly, the procedure for a Southern blot is as follows. Genomic DNA is isolated from a sample obtained from the patient. One can obtain DNA from almost any cellular tissue of the patient. The DNA is digested using one or more restriction enzymes, after which it is size-
30 fractionated by electrophoresis through an agarose slab gel. The DNA is then immobilized by transfer from the gel to a membrane (commonly nylon or nitrocellulose).

If all or part of the DPD gene is missing from the patient's genomic DNA, the probe will not hybridize to the genomic DNA, or else will hybridize to a

different-sized restriction fragment compared to the wild-type DPD gene. If a patient is heterozygous at the DPD locus, the clinician will observe either a reduced hybridization signal compared to wild-type (probe region deleted from one of the two alleles) or hybridization to two different-sized restriction fragments (part of one DPD gene deleted). If a sample from a patient lacks a gene that codes for DPD, the clinician should exercise extreme caution in using 5-FU as chemotherapy. A patient who is missing all or part of one or both DPD genes (*e.g.*, either a heterozygote or homozygote for a defective DPD gene) is at risk of 5-FU toxicity or conditions such as thymine uraciluria that are due to inadequate levels of DPD activity.

DPD deficiency that results in 5-FU toxicity or thymine uraciluria might also result from insufficient DPD mRNA levels. The Northern blot is a particularly useful method for detecting DPD mRNA levels. By detecting DPD mRNA levels, rather than detecting the presence of the DPD gene, Northern blots permit quantitation of DPD gene expression. This facilitates identification of patients who are DPD deficient for any of several reasons. A homozygote in which both DPD alleles are deleted will produce no DPD mRNA, while a heterozygote will generally have an intermediate level of DPD mRNA compared to a patient who is homozygous wild type. A Northern blot also allows the clinician to identify patients who, although they carry DPD genes, have a lower than normal level of DPD gene expression. Such patients are also at risk of 5-FU toxicity and thymine uraciluria.

Suitable samples for detection of DPD mRNA include any cells from the patient that express the DPD gene. Preferably, the cells will be obtained from a tissue that has high levels of DPD activity. In humans, the liver and lymphocytes generally have the highest DPD activity, with other tissues having less activity [Naguib *et al.* (1985) *Cancer Res.* 45: 5405-5412]. Because lymphocytes are much easier to isolate from a patient than liver cells, lymphocytes are a preferred sample for detecting DPD mRNA according to the claimed invention. However, one can also detect DPD mRNA in other cell types, such as fibroblasts.

Suitable methods for Northern blots are described in, for example, Sambrook, *supra.* and Chomczynski and Sacchi (1987) *Anal. Biochem.* 162: 156-159. Briefly, RNA is isolated from a cell sample using an extraction solution that releases the RNA from the cells while preventing degradation of the RNA. A commonly-used extraction solution contains a guanidinium salt. The RNA is purified from the extraction solution, such as by phenol-chloroform extraction followed by

ethanol precipitation. Optionally, one can separate the mRNA from ribosomal RNA and transfer RNA by oligo-dT cellulose chromatography, although such purification is not required to practice the claimed invention. The RNA is then size-fractionated by electrophoresis, after which the RNA is transferred from the gel to a nitrocellulose or nylon membrane. Labeled probes are used to ascertain the presence or absence of DPD-encoding mRNA.

If a sample from a patient has an insufficient amount of DPD nucleic acids, the patient is at risk of a toxic reaction to 5-FU, or is likely to suffer from thymine uraciluria or a related condition. Generally, an insufficient amount of DPD nucleic acids is less than about 70% of the normal amount of DPD nucleic acid, where "normal" refers to the amount of DPD nucleic acid found in the same amount of DNA or RNA from a sample that is not known to have a DPD deficiency. More typically, an amount of DPD that is less than about 50% of normal is indicative of an enhanced risk of 5-FU toxicity or thymine uraciluria.

Yet another potential cause of DPD deficiency in a patient is a missense or nonsense mutation in the DPD gene, or a mutation that interferes with mRNA processing. Our invention allows the clinician to detect these mutations. By choosing a probe that hybridizes to a mutant DPD gene, but not to the wild-type DPD gene (or vice versa), one can determine whether the patient carries an abnormal DPD gene that may result in inadequate expression of the DPD gene, or expression of an abnormal DPD enzyme that has less activity than the wild-type enzyme.

A variety of nucleic acid hybridization formats in addition to Northern and Southern blots are known to those skilled in the art. For example, common formats include sandwich assays and competition or displacement assays. Hybridization techniques are generally described in "*Nucleic Acid Hybridization, A Practical Approach*," Hames, B.D. and Higgins, S.J. (eds.), IRL Press, 1985; Gall and Pardue (1969) *Proc. Natl. Acad. Sci. USA*. 63: 378-383; and John *et al.* (1969) *Nature* 223: 582-587. These assays are sometimes preferred over classical Northern and Southern blots because of their greater speed and simplicity.

Sandwich assays are commercially useful hybridization assays for detecting or isolating nucleic acid sequences. These assays are easily automated, which results in a more cost-effective and sometimes more accurate assay. Sandwich assays utilize a "capture" nucleic acid that is covalently linked to a solid

support, and a labelled "signal" nucleic acid that is in solution. The clinical sample provides the target nucleic acid. The "capture" nucleic acid and "signal" nucleic acid probe each hybridize to the target nucleic acid to form a "sandwich" hybridization complex. To be effective, the signal nucleic acid cannot hybridize to the capture nucleic acid.

One embodiment of this invention embraces a kit that utilizes the concept of the sandwich assay. This kit includes a first component for the collection of samples from patients, vials for containment, and buffers for the dispersement and lysis of the sample. A second component contains media in either dry or liquid form for the hybridization of target and probe polynucleotides, as well as for the removal of undesirable and nonduplexed forms by washing. A third component includes a solid support upon which is fixed or to which is conjugated unlabeled nucleic acid probe(s) that is(are) complementary to a DPD nucleic acid. In the case of multiple target analysis more than one capture probe, each specific for its own DPD nucleic acid target region, will be applied to different discrete regions of the dipstick. A fourth component contains labeled probe that is complementary to a second and different region of the same DPD nucleic acid strand to which the immobilized, unlabeled nucleic acid probe of the third component is hybridized.

No matter which assay format is employed, labelled signal nucleic acids are typically used to detect hybridization. Complementary nucleic acids or signal nucleic acids can be labelled by any one of several methods typically used to detect the presence of hybridized polynucleotides, as described above. The most common method of detection is the use of autoradiography with ^3H , ^{125}I , ^{35}S , ^{14}C , or ^{32}P -labelled probes or the like. Other labels include ligands which bind to labelled antibodies, fluorophores, chemiluminescent agents, enzymes, and antibodies which can serve as specific binding pair members for a labelled ligand.

Detection of a hybridization complex may require the binding of a signal generating complex to a duplex of target and probe polynucleotides or nucleic acids. Typically, such binding occurs through ligand and anti-ligand interactions as between a ligand-conjugated probe and an anti-ligand conjugated with a signal. The label can also allow indirect detection of the hybridization complex. For example, where the label is a hapten or antigen, the sample can be detected by using antibodies. In these systems, a signal is generated by attaching fluorescent or enzyme molecules to the antibodies or, in some cases, by attachment to a

radioactive label. [Tijssen, P., "Practice and Theory of Enzyme Immunoassays," *Laboratory Techniques in Biochemistry and Molecular Biology*, Burdon, R.H., van Knippenberg, P.H., Eds., Elsevier (1985), pp. 9-20].

5 The sensitivity of the hybridization assays can be enhanced through use of a nucleic acid amplification system that multiplies the target nucleic acid being detected. Examples of such systems include the polymerase chain reaction (PCR) system and the ligase chain reaction (LCR) system. Other methods recently described in the art are the nucleic acid sequence based amplification (NASBA", Cangene, Mississauga, Ontario) and Q Beta Replicase systems. Amplification
10 methods permit one to detect the presence or absence of DPD nucleic acids using only a very small sample. Furthermore, amplification methods are especially amenable to automation.

 One preferred method for detecting DPD deficiency is reverse transcriptase PCR (RT-PCR). Briefly, this method involves extracting RNA from th
15 sample being analyzed, making a cDNA copy of the mRNA using an oligo-dT primer and reverse transcriptase, and finally amplifying part or all of the cDNA by PCR. For primers, one can use oligonucleotide primers that are complementary to the 5' and 3' sequences that flank the DNA region to be amplified. One can select primers to amplify the entire region that codes for a full-length DPD polypeptide, or
20 to amplify smaller DNA segments that code for part of the DPD polypeptide, as desired. For human DPD analysis, suitable pairs of primers include: SEQ. ID Nos. 5 and 6, SEQ. ID Nos. 7 and 8, and SEQ. ID Nos. 9 and 10. A detailed example of RT-PCR analysis as used for detection of DPD deficiency is presented in Example 4 below.

25 An alternative means for determining the level at which a DPD gene is expressed is *in situ* hybridization. *In situ* hybridization assays are well known and are generally described in Angerer *et al.* (1987) *Methods Enzymol.* 152: 649-660. In an *in situ* hybridization assay, cells are fixed to a solid support, typically a glass slide. If DNA is to be probed, the cells are denatured with heat or alkali. The c lls
30 are then contacted with a hybridization solution at a moderate temperature to permit annealing of labeled probes specific to DPD-encoding nucleic acids. The probes are preferably labelled with radioisotopes or fluorescent labels.

C. Expression of Recombinant Dihydropyrimidine Dehydrogenase

The present invention also provides methods for expressing recombinant dihydropyrimidine dehydrogenase (DPD). These methods involve cloning the claimed isolated DPD cDNA into an appropriate expression vector, transforming the expression vector into a host cell, and growing the host cells under conditions that lead to expression of the DPD cDNA. Numerous expression systems are suitable for expression of cDNA encoding DPD. Because these basic techniques are known to those of skill in the art, no attempt is made here to describe in detail the various basic methods known for the expression of proteins in prokaryotes or eukaryotes.

In brief summary, the expression of natural or synthetic nucleic acids encoding DPD will typically be achieved by operably linking a DPD-encoding cDNA to a promoter that functions in the host cell of choice. Either constitutive or inducible promoters are suitable. This "expression cassette" is typically incorporated in an expression vector. The vectors contain regulatory regions that cause the vector to replicate autonomously in the host cell, or else the vector can replicate by becoming integrated into the genomic DNA of the host cell. Suitable vectors for both prokaryotes and eukaryotes are known to those of skill in the art. Typical expression vectors can also contain transcription and translation terminators, translation initiation sequences, and enhancers that are useful for regulating the amount of DPD expression. To obtain high level expression of a cloned gene, such as those polynucleotide sequences encoding DPD, it is desirable to construct expression vectors that contain, at minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/ translation terminator. Expression vectors often contain control elements that permit the vector to replicate in both eukaryotes and prokaryotes, as well as selectable markers that function in each. See, e.g., Sambrook, *supra.*, for examples of suitable expression vectors.

1. Expression in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines, bird, fish, and mammalian cells, are known to those of skill in the art. Eukaryotic systems, including yeast, mammalian, and insect, suitable for expressing
5 DPD are discussed briefly below.

Synthesis of heterologous proteins in yeast is well known. *Methods in Yeast Genetics*, Sherman, F., et al., Cold Spring Harbor Laboratory, (1982) is a well recognized work describing the various methods available to produce the protein in yeast. Suitable vectors for expression in yeast usually have expression
10 control sequences, such as promoters, including 3-phosphoglycerate kinase or other glycolytic enzymes, and an origin of replication, termination sequences and the like as desired. For instance, suitable vectors are described in the literature (Botstein, et al., 1979, *Gene*, 8:17-24; Broach, et al., 1979, *Gene*, 8:121-133). Several commercial manufacturers of molecular biology reagents sell expression vectors
15 that are suitable for use in different eukaryotic host cells [See, e.g., product catalogs from Stratagene Cloning Systems, La Jolla Ca; Clontech Laboratories, Palo Alto CA; Promega Corporation, Madison WI]. These vectors are used as directed by the manufacturers except for the modifications described below that are necessary for expression of DPD.

20 Two procedures are commonly used to transform yeast cells. The first method involves converting yeast cells into protoplasts using an enzyme such as zymolyase, lyticase or glucanase. The protoplasts are then exposed to DNA and polyethylene glycol (PEG), after which the PEG-treated protoplasts are then regenerated in a 3% agar medium under selective conditions. Details of this
25 procedure are given in the papers by Beggs (1978) *Nature* (London) 275: 104-109 and Hinnen et al. (1978) *Proc. Natl. Acad. Sci. USA* 75: 1929-1933. The second procedure does not involve removal of the cell wall. Instead the cells are treated with lithium chloride or acetate and PEG and put on selective plates [Ito et al. (1983) *J. Bact.* 153: 163-168].

30 The DPD polypeptides, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques, or radioimmunoassay or other standard immunoassay techniques.

Higher eukaryotes are also suitable host cells for expression of recombinant DPD. Again, previously described methods are suitable, except that the modifications described below are necessary for efficient expression of DPD. Expression vectors for use in transforming, for example, mammalian, insect, bird, and fish cells are known to those of skill in the art.

Mammalian cells are illustrative of the techniques used for expression of DPD in eukaryotic cells. Mammalian cells typically grow in the form of monolayers of cells, although mammalian cell suspensions may also be used. A number of suitable host cell lines capable of expressing intact proteins have been developed in the art, and include the HEK293, BHK21, and CHO cell lines, and various human cells such as COS cell lines, HeLa cells, myeloma cell lines, Jurkat cells, etc. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (e.g., the CMV promoter, a HSV *tk* (thymidine kinase) promoter or *pgk* (phosphoglycerate kinase) promoter), an enhancer [Queen *et al.* (1986) *Immunol. Rev.* 89:49], and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites (e.g., an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells useful for production of recombinant DPD are available, for instance, from the American Type Culture Collection Catalogue of Cell Lines and Hybridomas (7th edition, 1992), as well as from various commercial manufacturers of molecular biology reagents.

Insect cells are another eukaryotic system that is useful for expressing recombinant DPD protein. Appropriate vectors for expressing recombinant DPD in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth and *Drosophila* cell lines such as a Schneider cell line [See, Schneider J. (1987) *Embryol. Exp. Morphol.* 27:353-365].

Higher eukaryotic host cells, such as mammalian and insect cells, are rendered competent for transformation by various means. There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation and micro-injection of the DNA directly into the cells.

The transformed cells are cultured by means well known in the art. *Biochemical Methods in Cell Culture and Virology*, Kuchler, R.J., Dowden, Hutchinson and Ross, Inc (1977). The expressed polypeptides are isolated from cells grown as suspensions or as monolayers. The DPD polypeptides are recovered
5 by well known mechanical, chemical or enzymatic means.

2. Expression in Prokaryotes

A variety of prokaryotic expression systems can be used to express recombinant DPD. Examples of suitable host cells include *E. coli*, *Bacillus*,
10 *Streptomyces*, and the like. For each host cell, one employs an expression plasmids that contains appropriate signals that direct transcription and translation in the chosen host organism. Such signals typically include a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator. Examples of regulatory regions suitable for this
15 purpose in *E. coli* are the promoter and operator region of the *E. coli* tryptophan biosynthetic pathway as described by Yanofsky, C. (1984) *J. Bacteriol.* 158: 1018-1024 and the leftward promoter of phage lambda (λ) as described by Herskowitz and Hagen (1980) *Ann. Rev. Genet.* 14: 399-445. Several commercial manufacturers of molecular biology reagents sell prokaryotic expression vectors
20 that have been optimized for high levels of heterologous gene expression [See, e.g., product catalogs from Stratagene Cloning Systems, La Jolla Ca; Clontech Laboratories, Palo Alto CA; Promega Corporation, Madison WI]. These vectors are especially suitable for producing recombinant DPD, and are used as directed by the manufacturer, except that modifications to the growth medium are required for DPD
25 expression, as described below.

Suitable expression vectors for use in prokaryotes typically contain a selectable marker that, when cells are grown under appropriate conditions, cause only those cells that contain the expression vector to grow. Examples of such markers useful in *E. coli* include genes specifying resistance to ampicillin,
30 tetracycline, or chloramphenicol. See, e.g., Sambrook, *supra*. for details concerning selectable markers suitable for use in *E. coli*.

Overexpression of DPD causes elimination of pyrimidines from cells. This results in selection against cells that produce high levels of DPD. The present invention provides methods to circumvent this problem. These methods involve

adding uracil to the growth medium. Addition of other cofactors such as FAD and FMN also has a beneficial effect, although not as great as for uracil addition. For expression of DPD in *E. coli*, for example, a preferred medium is Terrific Broth [Tartof and Hobbs (1987) Bethesda Research Labs FOCUS 9: 12] that contains 100 $\mu\text{g/ml}$ ampicillin or other antibiotic suitable for the selectable marker contained on the expression vector employed. To allow growth of cells that express DPD, the medium is typically supplemented with 100 μM uracil, and optionally 100 μM each of FAD and FMN, and 10 μM each of $\text{Fe}(\text{NH}_4)_2\text{SO}_4$ and Na_2S .

Recombinant DPD produced by prokaryotic cells may not necessarily fold into the same configuration as eukaryotically-produced DPD. If improper folding inhibits DPD activity, one can "refold" the DPD polypeptide by first denaturing the protein, and then allowing the protein to renature. This can be accomplished by solubilizing the bacterially produced proteins in a chaotropic agent such as guanidine HCl, reducing all the cysteine residues by using a reducing agent such as β -mercaptoethanol. The protein is then renatured, either by slow dialysis or by gel filtration. See, e.g., U.S. Patent No. 4,511,503.

Detection of the expressed antigen is achieved by methods known in the art as radioimmunoassay, or Western blotting techniques or immunoprecipitation. Purification from *E. coli* can be achieved following procedures described in, for example, U.S. Patent No. 4,511,503.

3. Purification of DPD Polypeptides

The DPD polypeptides produced by recombinant DNA technology as described herein can be purified by standard techniques well known to those of skill in the art. Typically, the cells are lysed (e.g., by sonication) and the protein is then purified to substantial purity using standard techniques such as selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, e.g., R. Scopes, *Protein Purification: Principles and Practice*, Springer-Verlag: New York (1982), which is incorporated herein by reference. For example, one can raise antibodies against the DPD polypeptides and use the antibodies for immunoprecipitation or affinity chromatography using standard methods.

If the DPD polypeptide is produced as a fusion protein, in which the DPD moiety is fused to non-DPD amino acids, the desired polypeptide can be released by digestion with an appropriate proteolytic enzyme.

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D. Use of DPD nucleic acids as selectable markers

Another aspect of the claimed invention is the use of a DPD nucleic acid as a selectable marker that is effective in both prokaryotes and eukaryotes. Selectable markers are genes that, when present in a cloning vector, produce a gene product that enables cells containing the vector to grow under conditions that prevent cells lacking the vector from growing. In contrast to the selectable markers of the invention, most selectable markers function only in one or the other of eukaryotes and prokaryotes, not in both. Thus, cloning vectors that are intended for propagation in both types of organisms usually require two different selectable markers.

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The claimed selectable markers are DPD-encoding nucleic acids. Cells that express these nucleic acids are resistant to 5-FU. 5-fluorouracil, which is toxic to both prokaryotic and eukaryotic cells, is degradatively inactivated by DPD. Therefore, one can select cells that contain a DPD nucleic acid that is operably linked to a promoter simply by growing the cells in the presence of 5-FU. To practice the invention, one operably links the DPD nucleic acid to a promoter that functions in the host cell of interest. Suitable promoters and other control signals are described above. In a preferred embodiment, the DPD nucleic acid is integrated into an expression cassette that functions in both prokaryotes and eukaryotes. One example of such a bifunctional expression cassette is the ZAP Express™ expression cassette (Stratagene, La Jolla CA), which is described in U.S. Patent No. 5,128,256. The DPD nucleic acid is inserted into the multiple cloning site which is downstream of a tandem array that includes both prokaryotic and eukaryotic transcription and translation regulatory sequences.

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To determine appropriate growth conditions for using the DPD selectable marker, one first tests the untransformed host cells of interest for ability to grow in medium containing various amounts of 5-FU. A 5-FU concentration that results in complete or nearly complete inhibition of host cell growth is then employed in the medium used to select transformants. The amount of 5-FU

required may vary depending on the particular medium used, the host cells, and whether the cells are grown in liquid culture or on a solid medium such as agar.

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EXAMPLES

Example 1: Cloning and Characterization of Pig and Human DPD cDNAs

In this Example, we describe the cloning and characterization of cDNAs for pig and human dihydropyrimidine dehydrogenases.

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MATERIALS AND METHODS

We isolated total RNA from frozen pig liver using the method of Chirgwin *et al.* (1979) *Biochemistry* 18: 5294-5299, except that we used CsTFA (Pharmacia, Inc., Milwaukee, WI) instead of CsCl. We extracted the RNA twice with phenol-chloroform emulsion and then ethanol precipitated the RNA prior to use. Next, we isolated poly(A) RNA by oligo (dT)-cellulose chromatography [Aviv and Leder (1977) *Proc. Nat'l. Acad. Sci. USA* 69: 1408-1412] and used it as a template for synthesis of cDNA. We used oligo-dT as a primer, and extended the primer using reverse transcriptase. Then, we made the cDNA double-stranded and cloned it into λ gt24A using a kit supplied by Gibco BRL Life Technologies, Inc., Gaithersburg, MD. The DNA was packaged using the λ packaging system from Gibco BRL. We plated the phage particles in *Escherichia coli* Y1090r.

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To identify plaques that express pig DPD, we screened the library using a polyclonal antibody against pig DPD [Podschun *et al.* (1989) *Eur. J. Biochem.* 185: 219-224]. We obtained a partial cDNA that we used to rescreen the library in *E. coli* Y1088 by plaque hybridization. This yielded a cDNA that contained the complete DPD reading frame. We subcloned the cDNA into the *Not*I and *Sal*I sites of the plasmid vector pSport (Gibco BRL).

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To clone the human DPD cDNA, we used a fragment of the pig cDNA that includes most of the coding region to screen previously amplified human liver cDNA libraries that had been prepared in λ gt11 [Yamano *et al.* (1989) *Biochemistry* 28: 7340-7348]. We isolated the human DPD cDNA as three overlapping fragments, which we subcloned into the *Eco* RI site of pUC18. The three fragments were joined together using overlapping *Cla* I sites in pUC18. We then determined the complete sequences of pig and human DPD cDNAs using an Applied

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Biosystems 373A DNA sequencer, synthetic primers, and fluorescent dye terminator chemistry as described by the manufacturer. The oligonucleotide primers were synthesized using a CENTRICON 10™ filter (Millipore Corp.). Each base was determined at least once on both strands. The DNA and deduced amino acid sequences were analyzed using MacVector sequence analysis software (International Biotechnologies, Inc., New Haven, CT).

RESULTS

We isolated partial pig cDNAs by screening 1×10^6 plaques from an unamplified λ gt22A library. After verification by sequencing, we used a partial cDNA to rescreen 500,000 plaques. Four cDNAs were isolated which contained inserts of about 4.5 kb. We completely sequenced one of these and found that it encompassed the full coding region of the protein (Figures 2A-2B). The deduced amino acid sequence of the amino terminal region agrees with the amino acid sequence determined from the pig enzyme [Podschun *et al.* (1989) *Eur. J. Biochem.* 185: 219-224. A number of segments of amino acids previously sequenced were found in the cDNA-deduced amino acid sequence (Figure 3, underlined). These were determined by cyanogen bromide cleavage (residues 117-127) and trypsin cleavage (residues 260-277; 308-315; 656-682; 904-913) followed by HPLC separation and sequencing (data not shown). The first residue of the amino terminal portion of the 12,000 dalton cleavage fragment from the pig DPD is shown by a vertical arrow at residue 904. These data establish the pig DPD open reading frame of 1025 amino acids.

The nucleotide sequence of the human DPD is shown in Figures 1A-1B. The deduced amino acid sequence of the human DPD is identical to that of the pig DPD, except where indicated in Figure 3. The calculated molecular weights are 11,416 and 111,398 daltons for pig and human DPD, respectively. The poly(A) addition sequence of AAATAAA is found 17 bp upstream of a putative poly(A) tract cloned in the cDNA. This 3'-untranslated region was not isolated in the human cDNA clones.

The cDNA-derived protein sequences revealed the presence of a number of putative binding sites for known DPD cofactors. Recent EPR measurements on DPD from *Alcaligenes eutrophus* confirmed the existence of FMN, iron, and acid-labile sulfide, the latter two of which are indicative of iron sulfur

clusters [Schmitt *et al.* (1994) *J. Inorg. Biochem.* (in press). The C-terminal 12 kDa peptide fragment purified from the pig DPD shows absorbance in the 500-600 nm region and contains eight iron and eight acid-labile sulfides (Podschun *et al.* (1989), *supra.*). The binding site of iron-sulfur clusters contain Cys residues, a large number of which are found in the N-terminal half of the protein. However, these do not exhibit the typical motif pattern seen in other well-characterized iron sulfur-containing proteins. In the C-terminal region of pig and human DPD are typical motifs CXXCXXCXXXCX (SEQ ID No. 11) and CXXCXXCXXXCPC (SEQ ID No. 12) for [4Fe-4S] clusters [Dupuis *et al.* (1991) *Biochemistry* 30: 2954-2960] between residues 953 and 964 and residues 986 and 997, respectively. These lie within the 12 kDa iron-sulfur cluster-containing peptide [Podschun *et al.* (1989), *supra.*). No other [4Fe-4S] clusters were detected; however, other types of iron sulfur clusters such as [2Fe-2S] might be possible.

A typical NADPH binding motif VXVXGXGXGXXXAXXA (SEQ ID No. 13) [Wierenga *et al.* (1985) *Biochemistry* 24: 1346-1357] begins with V-335, except that the Gly at position 10 is an Ala in pig and human DPD. A motif for FAD binding, TXXXVFXGD [Eggink *et al.* (1990) *J. Mol. Biol.* 212: 135-142], is in the N-terminal region starting with T-471 and ending with D-481.

We elucidated the putative uracil binding site of DPD by incubating DPD in the presence of 5-iodouracil, a suicide inactivator of the bovine enzyme, and sequencing the modified chymotryptic peptide [Porter *et al.* (1991) *J. Biol. Chem.* 266: 19988-19994]. The corresponding sequence obtained is located between G-661 and R-678 in the primary protein sequence. Thus, the order of the functional domains of DPD is, from the N-terminus, NADPH/NADP-FAD-uracil-[4Fe-4S].

Example 2: Chromosome localization of the DPD gene

We localized the DPD gene to a specific human chromosome using a somatic cell hybrid strategy. Human-mouse and human-hamster cell lines were generated and characterized as described by McBride *et al.* [(1982a) *Nucl. Acids Res.* 10: 8155-8170; (1982b) *J. Exp. Med.* 155: 1480-1490; (1982c) *Proc. Nat'l. Acad. Sci. USA* 83: 130-134]. The human chromosome of each cell line was determined by standard isoenzyme analyses as well as by Southern analysis with probes from previously localized genes, and frequently, by cytogenetic analysis. Southern blots of hybrid cell DNA restriction digests on positively charged nylon

membranes were prepared after (0.7%) agarose gel electrophoresis and hybridized at high stringency with ^{32}P -labeled probes under conditions allowing no more than 10% divergence of hybridizing sequences.

We localized the DPD gene to human chromosome 1 by Southern analysis of a panel of human/rodent somatic cell hybrid DNAs digested with *Eco* RI using a 3' coding cDNA fragment as probe (Table 1). The gene segregated discordantly ($\geq 14\%$) with all other human chromosomes. The 3' probe identified a series of bands in human DNAs ranging in size from 0.8 to 1.5 kb. All hybridizing human bands appeared to cosegregate indicating that these bands were all present on the same chromosome. We then sub-localized the gene on chromosome 1 by analysis of hybrids containing spontaneous breaks and translocations involving this chromosome. One human/hamster hybrid with a break between NRAS (1p12) and PGM1 (1p22) retained the telomeric portion of the chromosome 1 short arm but the DPD gene was absent from this hybrid. Another human/hamster hybrid and a human/mouse hybrid each retained all, or nearly all, of the short arm of chromosome 1 including NRAS and all other short arm markers but all long arm markers were absent including a cluster of genes at 1q21 (trichohyalin, loricrin, and filaggrin); the human DPD gene was present in both of these hybrids. Finally, one additional human/hamster hybrid retained a centromeric fragment of chromosome 1 with the breakpoints on the long arm and short arm proximal to 1q21 and proximal to 1p31, respectively, and human DPD was present in this hybrid. These results indicate that the DPD gene can be sublocalized to the region 1p22-q21.

We confirmed these results by Southern analysis of the same panel of hybrids with a DPD 5' cDNA probe which detected 1.5, 5.0, 8.7, and 11.6 kb bands in human *Eco*RI digests. Both probes were used to examine DNAs from ten unrelated individuals separately digested with 12 different restriction enzymes for RFLPs. However, no polymorphisms were detected. A large number of hybridizing bands were detected with both DPD probes and these bands cosegregated indicating that they are all localized to the centromeric region of human chromosome 1 (i.e., 1p22-q21). A number of cross-hybridizing hamster and mouse bands were also identified with these probes. These results are consistent with the interpretation that there may be a single reasonably large gene (spanning at least 80 kb) in each of these species, and all hybridizing bands arise from a single gene.

However, we currently cannot exclude the possibility that the many hybridizing bands arise from a cluster of tandemly linked genes.

Recently, the human DPD gene (named "DPYD" by the human gene nomenclature committee) was more precisely mapped to 1p22 [Takai *et al.* (1994) (submitted for publication)].

Example 3: Expression of Pig DPD in *E. coli*

In this Example, we demonstrate the heterologous expression of a DPD polypeptide in a prokaryotic organism. Because large amounts of DPD protein are toxic to the host cells under normal growth conditions, additional components such as uracil are required in the medium.

METHODS

Construction of the Expression Plasmid. We constructed an expression plasmid by subcloning the pig DPD cDNA into the vector pSE420 (Invitrogen Corp., San Diego, CA). The cDNA contains an *Nco* I site coincident with the start codon (CCATGG) which was joined to the *Nco* I site in the vector that is in frame with the bacterial initiator Met. The pig DPD cDNA was inserted into pSE420 as an *Nco*I/*Afl*III fragment from the pSPORT vector in which the pig DPD cDNA had previously been subcloned.

DPD Expression in Escherichia coli. For each expression experiment, a single colony from a freshly made transformation of DH-5 α cells with the expression vector was inoculated in LB broth and grown to stationary phase. An aliquot from this culture was used to inoculate 250 ml of terrific broth containing 100 μ g/ml ampicillin and supplemented with 100 μ M of each FAD and FMN, 100 μ M uracil and 10 μ M each of Fe(NH₄)₂(SO₄) and Na₂S. Following a 90 min incubation at 29°C, we induced the *trp-lac* promoter in the expression vector by the addition of 1 mM isopropyl- β -D-thiogalacto-pyranoside (IPTG) and the culture was incubated for an additional 48 h.

The cells were then sedimented, washed twice with 250 ml of phosphate buffered saline (PBS) and resuspended in 45 ml of 35 mM potassium phosphate buffer (pH 7.3) containing 20% glycerol, 10 mM EDTA, 1 mM DTT, 0.1 mM PMSF and 2 μ M leupeptin. The cell suspension was lysed at 4°C with four 30 sec bursts of a Heat Systems sonicator model W 225-R at 25% of full power (Heat

Systems-Ultrasonics, Inc., Plain View NY). The resultant lysate was centrifuged at 100,000 x g for 60 min at 4°C. We then slowly added solid (NH₄)₂SO₄ to the supernatant at 4°C with gentle stirring to give a final concentration of 30% saturation. The precipitate was sedimented and the pellet containing expressed DPD was resuspended in 5 ml of 35 mM potassium phosphate buffer (pH = 7.3) containing 1 mM EDTA/1 mM DTT and 0.1 mM PMSF. The protein solution was dialyzed at 4°C for 36 h against 3 changes of 4 liters each of buffer and stored at -70°C until further use.

Catalytic assay. DPD activity was determined at 37°C by measuring the decrease in absorbance at 340 nm associated with the oxidation of NADPH to NADP⁺. The reaction mixture contained 28 mM potassium phosphate buffer (pH 7.3), 2 mM MgCl₂, 1 mM DTT, 60 μM NADPH and the expressed DPD in a final volume of 1 ml. The measurements were carried out using an Aminco DW-2000 double beam spectrophotometer using a blank that contained the complete reaction mixture except substrate. The reactions were initiated by addition of substrate (uracil, 5-fluorouracil or thymine). The catalytic activity was calculated as μmole of NADPH oxidized per minute and per mg of expressed DPD. Protein quantities were determined using the bicinchronic (BCA) procedure from Pierce Chemical Co., Rockford, IL) following the manufacturer's directions.

Analysis of cDNA-Expressed DPD Protein. SDS-polyacrylamide gel electrophoresis was carried out following the method of Laemmli [(1970) *Nature* 227: 680-685] using 8% acrylamide slab gels. The SDS-page gels were transferred to a nitrocellulose membrane by electroblotting for 90 min at 1.5 mA/cm² [Towbin *et al.* (1979) *Proc. Nat'l Acad. Sci. USA* 76: 4350-4354]. The membranes were blocked at room temperature using phosphate buffered saline (PBS) containing 0.5% Tween 20 and 3% skim milk. After blocking, the membranes were incubated for 4 h at room temperature with rabbit anti pig DPD polyclonal antibody dilute 200-fold in PBS. The membranes were washed three times in PBS containing 0.5% Tween 20 and rinsed twice with PBS prior to addition of alkaline phosphatase-labeled goat anti-rabbit IgG. Incubation was continued for 90 min and the membranes were developed using the reagent BCIP/NBT (Kikegaard & Perry Labs. Gaithersburg, MD).

RESULTS

The pig DPD was expressed in bacteria using the vector pSE 420 which has a *trp-lac* promoter that is inducible by isopropyl- β -d-thiogalactopyranoside (IPTG). Optimal expression was obtained when cells were grown at a temperature between 26°C and 30°C. Growth at higher temperatures resulted in aggregation of the protein in inclusion bodies. A number of cofactors known to be associated with the enzyme were added to the medium; the most critical was uracil which resulted in a greater than five-fold increase in DPD expression levels, compared to cells grown in unsupplemented medium.

The recombinantly expressed DPD enzyme comigrated with the intact 102 kDa DPD purified from pig liver and reacted with rabbit polyclonal antibody [Podschun *et al.* (1989) *supra.*] directed against the pig enzyme. DPD protein was undetectable in cells containing the expression vector without the DPD cDNA insert. The DPD purified from pig liver frequently has a second higher mobility band of about 12 kDa that results from a protease-labile site that liberates the iron sulfur-containing C-terminal fragment [Podschun *et al.* (1989) *supra.*].

The bacterially-expressed enzyme is produced intact and could be significantly purified away from other *E. coli* proteins by a single ammonium sulfate fractionation. By use of the purified pig DPD as a standard, we estimate that 50 to 100 mg of DPD were produced per liter of *E. coli* culture.

We tested the recombinantly expressed DPD enzyme for ability to metabolize typical DPD substrates such as uracil, thymine and 5-fluorouracil. Kinetic studies revealed that the recombinant DPD follows the ping pong reaction mechanism as previously shown for purified pig DPD [Podschun *et al.* (1989), *supra.*]. The K_m 's of the recombinant DPD are of similar magnitude to the values published for the purified pig [Podschun *et al.* (1989), *supra.*], human [Lu *et al.* (1992) *J. Biol. Chem.* 267: 17102-17109] and rat DPD enzymes [Fujimoto *et al.* (1991) *J. Nutr. Sci. Vitaminol.* 37: 89-98]. The V_{max} values of expressed DPD were about three to five-fold lower than the purified pig enzyme reflecting the fact that the expressed DPD was only partially purified. However, these data establish that the expressed enzyme reflects the properties of the purified pig liver DPD. Thus, *E. coli* should prove useful for examining any enzymatic variants obtained

through screening DPD-deficient individuals and for preparing large amounts of intact holoenzyme for physico-chemical analysis.

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Example 4: Identification of mutations within *DPYD* gene

In an effort to understand the genetic basis for DPD deficiency, we analyzed a Dutch family that included a DPD-deficient individual. We determined the phenotype for thymine metabolism and related it to the DPD protein content in
10 fibroblasts. Then we identified the genetic defect using RT-PCR and found that the deficiency was due to a homozygous deletion in the DPD mRNA. The deleted portion corresponded to an exon in the *DPYD* gene. This phenotype/genotype relationship accounts for the DPD metabolic disorder in the patient. Additionally, we confirmed an autosomal recessive pattern of inheritance for DPD deficiency.

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METHODS

Isolation of RNA. RNA was isolated from cultures of human fibroblast corresponding to all five subjects used in this study by the guanidinium thiocyanate phenol-chloroform method [Chomczynski and Sacchi (1987) *Anal. Biochem.* 162:
20 156-159]. The RNA was dissolved in water and stored at -80°C until further use.

RT-PCR. cDNA was synthesized by reverse transcription from total RNA isolated from cultured fibroblast. About 1 µg of total RNA was mixed with oligo-dT primers and incubated at 65°C for 15 min to denature secondary structure
25 in the template. The primed RNA was incubated for 60 min at 40°C in 20 µl of a reaction mixture containing 100 mM Tris-HCl (pH 8.3), 40 mM KCl, 10 mM MgCl₂, 50 µM spermidine, 100 mM dNTPs, 4 mM sodium phosphate, 0.5 units placental RNase inhibitor and 0.5 units of AMV reverse transcriptase (Invitrogen, CA). The synthesis reaction was repeated once by the addition of 0.5 units of fresh revers
30 transcriptase. The cDNA was made double stranded by PCR without further purification. The coding region of the cDNA was amplified in three fragment with the primer pairs indicated in Table 1.

Table 1: Primer pairs for RT-PCR analysis of human DPD cDNA (hDPD).

5	Fragment amplified	Location in hDPD cDNA (nucleotides)	Primer sequence	SEQ. ID
				No.
10	1.5 kb	RTF1:36 - 55	5'GCAAGGAGGGTTTGTCACTG3'	5
		RTR1:1558 - 1536	5'CCGATTCCACTGTAGTGTAGCC3'	6
	906 bp	H13:1539 - 1558	5'TAACACTACAGTGGAAATCGG3'	7
		RTR4:2445 - 2426	5'AAATCCAGGCAGAGCACGAG3'	8
15	919 bp	RTR5:2424 - 2447	5'TGCTCGTGCTCTGCCTGGATTTC3'	9
		RTR5:3343 - 3320	5'ATTGAATGGTCATTGACATGAGAC3'	10

We carried out PCR in 50 μ l of a reaction mixture consisting of 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 2.5 mM MgCl₂, 0.5 mM dNTPs, 1 μ M primers and 2.5 units Taq polymerase (Perkin-Elmer Cetus). Thirty cycles were used, each cycle consisted of denaturing at 96°C for 1 min, annealing at 55°C for 1 min and extending at 72°C for 2 min. The amplified products were extracted with 1 volume chloroform and purified by filtration through Centricon™ 100 filter units (Amicon, Inc. Beverly WA). Typically, we used one fifth of the PCR product for DNA sequence analyses with an Applied Biosystems 373A automated sequencer and fluorescent dye-deoxy terminator chemistry. We elucidated appropriate primers for DNA sequencing from the DPD cDNA sequence disclosed herein and synthesized the primers using an Applied Biosystems 394 DNA & RNA synthesizer. Sequence data have been analyzed using MacVector™ sequence analysis software (International Biotechnologies).

PCR Product Analysis and Southern Blots. We analyzed the PCR fragments by electrophoresis through a 1% agarose gel in the presence of ethidium bromide. Prior to Southern blotting, the gels were depurinated by a 20 min incubation in 200 mM HCl, after which we denatured the DNA by a 20 min incubation in 0.5 M NaOH. The DNA was transferred to Gene Screen Plus™ membranes (New England Biolabs) overnight in 0.5 M NaOH as the transfer solution. We fixed the DNA by baking at 80°C, prehybridized at 65°C for 3 h in a solution containing 6X SSC, 1X Denhardt's reagent, 0.5 % sodium dodecyl

sulfate and 0.2 mg/ml sonicated salmon sperm DNA. We then hybridized overnight at 65°C in the same solution containing 1.5×10^6 cpm/ml of ^{32}P random priming labelled human DPD cDNA. After washing at 65°C for 20 min in 2 x SSC, 0.5% SDS and 45 min 0.1 x SSC, 0.5% SDS at 65°C, the
5 membranes were exposed to X-ray film (Eastman Kodak, Co.) at -80°C for 30 min.

Western Immunoblots. We carried out SDS-PAGE gel electrophoresis using the method of Laemmli (1970) *Nature* 227: 107-111.
10 The gels were transferred to nitrocellulose by semi-dry electroblotting for 90 min at 1.5 mA/cm². We detected DPD polypeptides using rabbit anti-pig DPD primary antibody and the enhanced chemiluminescence (ECL) detection method (Amersham Corp.), following the directions supplied by the manufacturer. Protein concentrations were determined using the bicinchronic acid procedure
15 (Pierce Chemical Co., Rockford, IL) using bovine serum albumin as standard.

Catalytic Activity. We measured DPD activity in human fibroblast extracts by HPLC using a modification of the method described by Tuchman *et al.* (1989) *Enzyme* 42: 15-24, using [^{14}C]-thymine as substrate.
20

RESULTS

Clinical evaluation. We have studied the genetic basis for the complete lack of DPD activity in one of the members of the pedigree shown in Figure 4. The patient (subject 4) was admitted to the hospital at the age of 25
25 months with bilateral microphthalmia, iris and choroidea coloboma, and nystagmus, in addition to a gradually increasing psychomotor retardation. However, no growth retardation or neurological abnormalities were detected. All other members of the pedigree were healthy and showed no abnormalities. The patient was diagnosed to have severe thymine-uraciluria. Skin biopsies
30 were taken in order to establish fibroblast cultures that were used in this study.

RT-PCR analysis of the DPD mRNA in cultured fibroblasts.
Fibroblast total RNA from every subject was subjected to RT-PCR. The PCR

products were hybridized with the [³²P]-labelled human DPD cDNA and the result is shown in Figure 5. The coding sequence of the DPD cDNA was fully amplified in three fragments that span 1500, 906 and 919 bp. All the fragments are present every subject, including the patient. The 1500 and 919 bp fragments were constant in all subjects. However, the 906 bp fragment was found in only certain subjects and was in linkage disequilibrium with a fragment of 741 bp. The latter was homozygous in the deficient patient and found together with the predicted normal size fragment in both parents. One sibling was heterozygous and another was homozygous for the normal allele. To confirm the possibility of a deletion in the mRNA-derived cDNA associated with the *DPYD* alleles of these subjects, we sequenced the PCR fragments using nested primers and found that the 741 originated from the 906 bp fragment by a deletion of 165 bp. A schematic representation showing the structure of both mRNAs is shown in Figure 6. Through partial sequencing of the *DPYD* gene, we found that the deletion present in the mRNA was coincident with a splicing site located in the genomic sequence of the *DPYD* gene that comprises a 165 bp exon. We have also found that the DNA corresponding to the deletion is present in the genomic DNA from the fibroblast cell lines since, as shown in Figure 7, the deleted cDNA sequence can be amplified by PCR from the genomic DNA in the patient, as well as from genomic DNA from other members of the family. These results indicate that the variant transcript is not the result of a large deletion containing the missing exon, but rather is the result of a mutation that causes incorrect splicing.

Catalytic activity and DPD protein content. DPD activities from the fibroblast cell lines were determined by HPLC (Table I). The maximum activity, 1 nmol h⁻¹ mg protein⁻¹, corresponds to subject 3 that was homozygous for the normal mRNA. The parents and another sibling (subjects 4, 5, and 2) present a lower value and the patient, subject 1, had background activity. It should be noted that the DPD activity obtained in human fibroblast is about 8-9 times lower than the equivalent activity in DPD from human lymphocytes.

To determine if the DPD protein content in our subjects follows a pattern similar to that of the catalytic activity, we measured fibroblast DPD protein by Western blots. DPD protein was not detectable in the patient, but was found in two other members of his family (subjects 2 and 4 in Figure 4) who were analyzed for comparison.

The catalytic activity pattern correlates with the DPD protein content for the different subjects. As expected, the patient with only background DPD activity in his fibroblast has no detectable DPD band in the Western blot when using an anti-pig DPD polyclonal antibody, suggesting a complete lack of DPD protein. It is interesting to note that even though the DPD protein is defective and does not accumulate in the cell, the DPD mRNA is present, indicating that the defective mRNA is not particularly unstable as compared to the mRNA encoding the active DPD protein.

In conclusion, this study established with certainty that thymine uraciluria is due to a mutation in the *DPYD* gene.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described can be used in the practice or testing of the present invention, the preferred methods and materials are now described. All publications and patent documents referenced in this application are incorporated by reference.

It is understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and scope of the appended claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

5 (i) APPLICANT: GONZALEZ, Frank J.
FERNANDEZ-SALGUERO, Pedro

(ii) TITLE OF INVENTION: CLONING AND EXPRESSION OF cDNA FOR HUMAN
DIHYDROPYRIMIDINE DEHYDROGENASE

10 (iii) NUMBER OF SEQUENCES: 13

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15 (A) ADDRESSEE: Townsend and Townsend Kourie and Crew
(B) STREET: Steuart Street Tower, One Market Plaza
(C) CITY: San Francisco
(D) STATE: California
(E) COUNTRY: US
20 (F) ZIP: 94105-1493

(v) COMPUTER READABLE FORM:

25 (A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

30 (A) APPLICATION NUMBER: US not yet designated
(B) FILING DATE: 09-SEP-1994
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

35 (A) NAME: Smith, Timothy L.
(B) REGISTRATION NUMBER: 35,367
(C) REFERENCE/DOCKET NUMBER: 15280-210

(ix) TELECOMMUNICATION INFORMATION:

40 (A) TELEPHONE: (415) 543-9600
(B) TELEFAX: (415) 543-5043

(2) INFORMATION FOR SEQ ID NO:1:

45 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3957 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

55 (A) NAME/KEY: CDS
(B) LOCATION: 88..3162

(ix) FEATURE:

60 (A) NAME/KEY: misc_feature
(B) LOCATION: 1..3957
(D) OTHER INFORMATION: /product= "Human DPD"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

65 AGACACGCTG TCACTTGGCT CTCTGGCTGG AGCTTGAGGA CGCAAGGAGG GTTTGTCACT 60

GGCAGACTCG AGACTGTAGG CACTGCC ATG GCC CCT GTG CTC AGT AAG GAC 111

Met Ala Pro Val Leu Ser Lys Asp

44

5	TCG GCG GAC ATC GAG AGT ATC CTG GCT TTA AAT CCT CGA ACA CAA ACT Ser Ala Asp Ile Glu Ser Ile Leu Ala Leu Asn Pro Arg Thr Gln Thr	159
10	CAT GCA ACT CTG TGT TCC ACT TCG GCC AAG AAA TTA GAC AAG AAA CAT His Ala Thr Leu Cys Ser Thr Ser Ala Lys Lys Leu Asp Lys Lys His	207
15	TGG AAA AGA AAT CCT GAT AAG AAC TGC TTT AAT TGT GAG AAG CTG GAG Trp Lys Arg Asn Pro Asp Lys Asn Cys Phe Asn Cys Glu Lys Leu Glu	255
20	AAT AAT TTT GAT GAC ATC AAG CAC ACG ACT CTT GGT GAG CGA GGA GCT Asn Asn Phe Asp Asp Ile Lys His Thr Thr Leu Gly Glu Arg Gly Ala	303
25	CTC CGA GAA GCA ATG AGA TGC CTG AAA TGT GCA GAT GCC CCG TGT CAG Leu Arg Glu Ala Met Arg Cys Leu Lys Cys Ala Asp Ala Pro Cys Gln	351
30	AAG AGC TGT CCA ACT AAT CTT GAT ATT AAA TCA TTC ATC ACA AGT ATT Lys Ser Cys Pro Thr Asn Leu Asp Ile Lys Ser Phe Ile Thr Ser Ile	399
35	GCA AAC AAG AAC TAT TAT GGA GCT GCT AAG ATG ATA TTT TCT GAC AAC Ala Asn Lys Asn Tyr Tyr Gly Ala Ala Lys Met Ile Phe Ser Asp Asn	447
40	CCA CTT GGT CTG ACT TGT GGA ATG GTA TGT CCA ACC TCT GAT CTA TGT Pro Leu Gly Leu Thr Cys Gly Met Val Cys Pro Thr Ser Asp Leu Cys	495
45	GTA GGT GGA TGC AAT TTA TAT GCC ACT GAA GAG GGA CCC ATT AAT ATT Val Gly Gly Cys Asn Leu Tyr Ala Thr Glu Glu Gly Pro Ile Asn Ile	543
50	GGT GGA TTG CAG CAA TTT GCT ACT GAG GTA TTC AAA GCA ATG AGT ATC Gly Gly Leu Gln Gln Phe Ala Thr Glu Val Phe Lys Ala Met Ser Ile	591
55	CCA CAG ATC AGA AAT CCT TCG CTG CCT CCC CCA GAA AAA ATG TCT GAA Pro Gln Ile Arg Asn Pro Ser Leu Pro Pro Pro Glu Lys Met Ser Glu	639
60	GCC TAT TCT GCA AAG ATT GCT CTT TTT GGT GCT GGG CCT GCA AGT ATA Ala Tyr Ser Ala Lys Ile Ala Leu Phe Gly Ala Gly Pro Ala Ser Ile	687
65	AGT TGT GCT TCC TTT TTG GCT CGA TTG GGG TAC TCT GAC ATC ACT ATA Ser Cys Ala Ser Phe Leu Ala Arg Leu Gly Tyr Ser Asp Ile Thr Ile	735
70	TTT GAA AAA CAA GAA TAT GTT GGT GGT TTA AGT ACT TCT GAA ATT CCT Phe Glu Lys Gln Glu Tyr Val Gly Gly Leu Ser Thr Ser Glu Ile Pro	783
75	CAG TTC CGG CTG CCG TAT GAT GTA GTG AAT TTT GAG ATT GAG CTA ATG Gln Phe Arg Leu Pro Tyr Asp Val Val Asn Phe Glu Ile Glu Leu Met	831
80	AAG GAC CTT GGT GTA AAG ATA ATT TGC GGT AAA AGC CTT TCA GTG AAT Lys Asp Leu Gly Val Lys Ile Ile Cys Gly Lys Ser Leu Ser Val Asn	879
85	GAA ATG ACT CTT AGC ACT TTG AAA GAA AAA GGC TAC AAA GCT GCT TTC Glu Met Thr Leu Ser Thr Leu Lys Glu Lys Gly Tyr Lys Ala Ala Phe	927

265	270	275	280	
ATT GGA ATA GGT TTG CCA GAA CCC AAT AAA GAT GCC ATC TTC CAA GGC Ile Gly Ile Gly Leu Pro Glu Pro Asn Lys Asp Ala Ile Phe Gln Gly 285 290 295	975			
CTG ACG CAG GAC CAG GGG TTT TAT ACA TCC AAA GAC TTT TTG CCA CTT Leu Thr Gln Asp Gln Gly Phe Tyr Thr Ser Lys Asp Phe Leu Pro Leu 300 305 310	1023			
GTA GCC AAA GGC AGT AAA GCA GGA ATG TGC GCC TGT CAC TCT CCA TTG Val Ala Lys Gly Ser Lys Ala Gly Met Cys Ala Cys His Ser Pro Leu 315 320 325	1071			
CCA TCG ATA CGG GGA GTC GTG ATT GTA CTT GGA GCT GGA GAC ACT GCC Pro Ser Ile Arg Gly Val Val Ile Val Leu Gly Ala Gly Asp Thr Ala 330 335 340	1119			
TTC GAC TGT GCA ACA TCT GCT CTA CGT TGT GGA GCT CGC CGA GTG TTC Phe Asp Cys Ala Thr Ser Ala Leu Arg Cys Gly Ala Arg Arg Val Phe 345 350 355 360	1167			
ATC GTC TTC AGA AAA GGC TTT GTT AAT ATA AGA GCT GTC CCT GAG GAG Ile Val Phe Arg Lys Gly Phe Val Asn Ile Arg Ala Val Pro Glu Glu 365 370 375	1215			
ATG GAG CTT GCT AAG GAA GAA AAG TGT GAA TTT CTG CCA TTC CTG TCC Met Glu Leu Ala Lys Glu Glu Lys Cys Glu Phe Leu Pro Phe Leu Ser 380 385 390	1263			
CCA CGG AAG GTT ATA GTA AAA GGT GGG AGA ATT GTT GCT ATG CAG TTT Pro Arg Lys Val Ile Val Lys Gly Arg Ile Val Ala Met Gln Phe 395 400 405	1311			
GTT CGG ACA GAG CAA GAT GAA ACT GGA AAA TGG AAT GAA GAT GAA GAT Val Arg Thr Glu Gln Asp Glu Thr Gly Lys Trp Asn Glu Asp Glu Asp 410 415 420	1359			
CAG ATG GTC CAT CTG AAA GCC GAT GTG GTC ATC AGT GCC TTT GGT TCA Gln Met Val His Leu Lys Ala Asp Val Val Ile Ser Ala Phe Gly Ser 425 430 435 440	1407			
GTT CTG AGT GAT CCT AAA GTA AAA GAA GCC TTG AGC CCT ATA AAA TTT Val Leu Ser Asp Pro Lys Val Lys Glu Ala Leu Ser Pro Ile Lys Phe 445 450 455	1455			
AAC AGA TGG GGT CTC CCA GAA GTA GAT CCA GAA ACT ATG CAA ACT AGT Asn Arg Trp Gly Leu Pro Glu Val Asp Pro Glu Thr Met Gln Thr Ser 460 465 470	1503			
GAA GCA TGG GTA TTT GCA GGT GGT GAT GTC GTT GGT TTG GCT AAC ACT Glu Ala Trp Val Phe Ala Gly Gly Asp Val Val Gly Leu Ala Asn Thr 475 480 485	1551			
ACA GTG GAA TCG GTG AAT GAT GGA AAG CAA GCT TCT TGG TAC ATT CAC Thr Val Glu Ser Val Asn Asp Gly Lys Gln Ala Ser Trp Tyr Ile His 490 495 500	1599			
AAA TAC GTA CAG TCA CAA TAT GGA GCT TCC GTT TCT GCC AAG CCT GAA Lys Tyr Val Gln Ser Gln Tyr Gly Ala Ser Val Ser Ala Lys Pro Glu 505 510 515 520	1647			
CTA CCC CTC TTT TAC ACT CCT ATT GAT CTG GTG GAC ATT AGT GTA GAA Leu Pro Leu Phe Tyr Thr Pro Ile Asp Leu Val Asp Ile Ser Val Glu 525 530 535	1695			
ATG GCC GGA TTG AAG TTT ATA AAT CCT TTT GGT CTT GCT AGC GCA ACT Met Ala Gly Leu Lys Phe Ile Asn Pro Phe Gly Leu Ala Ser Ala Thr 1743				

46

	540	545	550	
5	CCA GCC ACC AGC ACA TCA ATG ATT CGA AGA GCT TTT GAA GCT GGA TGG Pro Ala Thr Ser Thr Ser Met Ile Arg Arg Ala Phe Glu Ala Gly Trp 555 560 565	1791		
10	GGT TTT GCC CTC ACC AAA ACT TTC TCT CTT GAT AAG GAC ATT GTG ACA Gly Phe Ala Leu Thr Lys Thr Phe Ser Leu Asp Lys Asp Ile Val Thr 570 575 580	1839		
15	AAT GTT TCC CCC AGA ATC ATC CGG GGA ACC ACC TCT GGC CCC ATG TAT Asn Val Ser Pro Arg Ile Ile Arg Gly Thr Thr Ser Gly Pro Met Tyr 585 590 595 600	1887		
20	GGC CCT GGA CAA AGC TCC TTT CTG AAT ATT GAG CTC ATC AGT GAG AAA Gly Pro Gly Gln Ser Ser Phe Leu Asn Ile Glu Leu Ile Ser Glu Lys 605 610 615	1935		
25	ACG GCT GCA TAT TGG TGT CAA AGT GTC ACT GAA CTA AAG GCT GAC TTC Thr Ala Ala Tyr Trp Cys Gln Ser Val Thr Glu Leu Lys Ala Asp Phe 620 625 630	1983		
30	CCA GAC AAC ATT GTG ATT GCT AGC ATT ATG TGC AGT TAC AAT AAA AAT Pro Asp Asn Ile Val Ile Ala Ser Ile Met Cys Ser Tyr Asn Lys Asn 635 640 645	2031		
35	GAC TGG ACG GAA CTT GCC AAG AAG TCT GAG GAT TCT GGA GCA GAT GCC Asp Trp Thr Glu Leu Ala Lys Lys Ser Glu Asp Ser Gly Ala Asp Ala 650 655 660	2079		
40	CTG GAG TTA AAT TTA TCA TGT CCA CAT GGC ATG GGA GAA AGA GGA ATG Leu Glu Leu Asn Leu Ser Cys Pro His Gly Met Gly Glu Arg Gly Met 665 670 675 680	2127		
45	GGC CTG GCC TGT GGG CAG GAT CCA GAG CTG GTG CGG AAC ATC TGC CGC Gly Leu Ala Cys Gly Gln Asp Pro Glu Leu Val Arg Asn Ile Cys Arg 685 690 695	2175		
50	TGG GTT AGG CAA GCT GTT CAG ATT CCT TTT TTT GCC AAG CTG ACC CCA Trp Val Arg Gln Ala Val Gln Ile Pro Phe Phe Ala Lys Leu Thr Pro 700 705 710	2223		
55	AAT GTC ACT GAT ATT GTG AGC ATC GCA AGA GCT GCA AAG GAA GGT GGT Asn Val Thr Asp Ile Val Ser Ile Ala Arg Ala Ala Lys Glu Gly Gly 715 720 725	2271		
60	GCC AAT GGC GTT ACA GCC ACC AAC ACT GTC TCA GGT CTG ATG GGA TTA Ala Asn Gly Val Thr Ala Thr Asn Thr Val Ser Gly Leu Met Gly Leu 730 735 740	2319		
65	AAA TCT GAT GGC ACA CCT TGG CCA GCA GTG GGG ATT GCA AAG CGA ACT Lys Ser Asp Gly Thr Pro Trp Pro Ala Val Gly Ile Ala Lys Arg Thr 745 750 755 760	2367		
70	ACA TAT GGA GGA GTG TCT GGG ACA GCA ATC AGA CCT ATT GCT TTG AGA Thr Tyr Gly Gly Val Ser Gly Thr Ala Ile Arg Pro Ile Ala Leu Arg 765 770 775	2415		
75	GCT GTG ACC TCC ATT GCT CGT GCT CTG CCT GGA TTT CCC ATT TTG GCT Ala Val Thr Ser Ile Ala Arg Ala Leu Pro Gly Phe Pro Ile Leu Ala 780 785 790	2463		
80	ACT GGT GGA ATT GAC TCT GCT GAA AGT GGT CTT CAG TTT CTC CAT AGT Thr Gly Gly Ile Asp Ser Ala Glu Ser Gly Leu Gln Phe Leu His Ser 795 800 805	2511		
85	GGT GCT TCC CTC CTC CAG GTA TGC AGT GCC ATT CAG AAT CAG GAT TTC Gly Ala Ser Val Leu Gln Val Cys Ser Ala Ile Gln Asn Gln Asp Phe 810 815 820	2559		

47

	810	915	820	
5	ACT GTG ATC GAA GAC TAC TGC ACT GGC CTC AAA GCC CTG CTT TAT CTG Thr Val Ile Glu Asp Tyr Cys Thr Gly Leu Lys Ala Leu Leu Tyr Leu 825 830 835 840			2607
10	AAA AGC ATT GAA GAA CTA CAA GAC TGG GAT GGA CAG AGT CCA GCT ACT Lys Ser Ile Glu Glu Leu Gln Asp Trp Asp Gly Gln Ser Pro Ala Thr 845 850 855			2655
15	GTG AGT CAC CAG AAA GGG AAA CCA GTT CCA CGT ATA GCT GAA CTC ATG Val Ser His Gln Lys Gly Lys Pro Val Pro Arg Ile Ala Glu Leu Met 860 865 870			2703
20	GAC AAG AAA CTG CCA AGT TTT GGA CCT TAT CTG GAA CAG CGC AAG AAA Asp Lys Lys Leu Pro Ser Phe Gly Pro Tyr Leu Glu Gln Arg Lys Lys 875 880 885			2751
25	ATC ATA GCA GAA AAC AAG ATT AGA CTG AAA GAA CAA AAT GTA GCT TTT Ile Ile Ala Glu Asn Lys Ile Arg Leu Lys Glu Gln Asn Val Ala Phe 890 895 900			2799
30	TCA CCA CTT AAG AGA AGC TGT TTT ATC CCC AAA AGG CCT ATT CCT ACC Ser Pro Leu Lys Arg Ser Cys Phe Ile Pro Lys Arg Pro Ile Pro Thr 905 910 915 920			2847
35	ATC AAG GAT GTA ATA GGA AAA GCA CTG CAG TAC CTT GGA ACA TTT GGT Ile Lys Asp Val Ile Gly Lys Ala Leu Gln Tyr Leu Gly Thr Phe Gly 925 930 935			2895
40	GAA TTG AGC AAC GTA GAG CAA GTT GTG GCT ATG ATT GAT GAA GAA ATG Glu Leu Ser Asn Val Glu Gln Val Val Ala Met Ile Asp Glu Glu Met 940 945 950			2943
45	TGT ATC AAC TGT GGT AAA TGC TAC ATG ACC TGT AAT GAT TCT GGC TAC Cys Ile Asn Cys Gly Lys Cys Tyr Met Thr Cys Asn Asp Ser Gly Tyr 955 960 965			2991
50	CAG GCT ATA CAG TTT GAT CCA GAA ACC CAC CTG CCC ACC ATA ACC GAC Gln Ala Ile Gln Phe Asp Pro Glu Thr His Leu Pro Thr Ile Thr Asp 970 975 980			3039
55	ACT TGT ACA GGC TGT ACT CTG TGT CTC AGT GTT TGC CCT ATT GTC GAC Thr Cys Thr Gly Cys Thr Leu Cys Leu Ser Val Cys Pro Ile Val Asp 985 990 995 1000			3087
60	TGC ATC AAA ATG GTT TCC AGG ACA ACA CCT TAT GAA CCA AAG AGA GGC Cys Ile Lys Met Val Ser Arg Thr Thr Pro Tyr Glu Pro Lys Arg Gly 1005 1010 1015			3135
65	GTA CCC TTA TCT GTG AAT CCG GTG TGT TAAGGTGATT TGTGAAACAG Val Pro Leu Ser Val Asn Pro Val Cys 1020 1025			3182
70	TTGCTGTGAA CTTTCATGTC ACCTACATAT GCTGATCTCT TAAAATCATG ATCCTTGTGT TCAGCTCTTT CCAAATTAAA ACAAATATAC ATTTTCTAAA TAAAAATATG TAATTTCAAA ATACATTTGT AAGTGTAATAA AATGTCTCAT GTCAATGACC ATTCAATTAG TGGCATAAAA TAGAATAATT CTTTCTGAG GATAGTAGTT AAATAACTGT GTGGCAGTTA ATTGGATGTT CACTGCCAGT TGTCTTATGT GAAAAATTAA CTTTTTGTGT GGCAATTAGT GTGACAGTTT CCAAATTGCC CTATGCTGTG CTCCATATTT GATTTCTAAT TGTAAGTGAA ATTAAGCATT TTGAAACAAA GTACTCTTTA ACATACAAGA AAATGTATCC AAGGAAACAT TTTATCAATA			3242 3302 3362 3422 3482 3542 3602

48

AAAATTACCT TTAATTTTAA TGCTGTTTCT AAGAAAATGT AGTTAGCTCC ATAAAGTACA 3662
 AATGAAGAAA GTCAAAAATT ATTTGCTATG GCAGGATAAG AAAGCCTAAA ATTGAGTTTG 3722
 5 TGGACTTTAT TAAGTAAAAT CCGCTTCGCT GAAATTGCTT ATTTTGGGTG TTGGATAGAG 3782
 GATAGGGAGA ATATTTACTA ACTAAATACC ATTGACTACT CATGCGTGAG ATGGGTGTAC 3842
 10 AAACATCC TCTTTTAATG GCATTTCTCT TTAAACTATG TTCCTAACCA AATGAGATGA 3902
 TAGGATAGAT CCTGGTTACC ACTCTTTTAC TGTGCACATA TGGGCCCGG AATTC 3957

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1025 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

25 Met Ala Pro Val Leu Ser Lys Asp Ser Ala Asp Ile Glu Ser Ile Leu
 1 5 10 15
 30 Ala Leu Asn Pro Arg Thr Gln Thr His Ala Thr Leu Cys Ser Thr Ser
 20 25 30
 35 Ala Lys Lys Leu Asp Lys Lys His Trp Lys Arg Asn Pro Asp Lys Asn
 35 40 45
 40 Cys Phe Asn Cys Glu Lys Leu Glu Asn Asn Phe Asp Asp Ile Lys His
 50 55 60
 45 Thr Thr Leu Gly Glu Arg Gly Ala Leu Arg Glu Ala Met Arg Cys Leu
 65 70 75 80
 50 Lys Cys Ala Asp Ala Pro Cys Gln Lys Ser Cys Pro Thr Asn Leu Asp
 85 90 95
 55 Ile Lys Ser Phe Ile Thr Ser Ile Ala Asn Lys Asn Tyr Tyr Gly Ala
 100 105 110
 60 Ala Lys Met Ile Phe Ser Asp Asn Pro Leu Gly Leu Thr Cys Gly Met
 115 120 125
 65 Val Cys Pro Thr Ser Asp Leu Cys Val Gly Gly Cys Asn Leu Tyr Ala
 130 135 140
 70 Thr Glu Glu Gly Pro Ile Asn Ile Gly Gly Leu Gln Gln Phe Ala Thr
 145 150 155 160
 75 Glu Val Phe Lys Ala Met Ser Ile Pro Gln Ile Arg Asn Pro Ser Leu
 165 170 175
 80 Pro Pro Pro Glu Lys Met Ser Glu Ala Tyr Ser Ala Lys Ile Ala Leu
 180 185 190
 85 Phe Gly Ala Gly Pro Ala Ser Ile Ser Cys Ala Ser Phe Leu Ala Arg
 195 200 205
 90 Leu Gly Tyr Ser Asp Ile Thr Ile Phe Glu Lys Gln Glu Tyr Val Gly
 210 215 220
 95 Gly Leu Ser Thr Ser Glu Ile Pro Gln Phe Arg Leu Pro Tyr Asp Val
 225 230 235 240

48

AAAATTACCT TTAATTTTAA TGCTGTTTCT AAGAAAATGT AGTTAGCTCC ATAAAGTACA 3662
 AATGAAGAAA GTCAAAAATT ATTTGCTATG GCAGGATAAG AAAGCCTAAA ATTGAGTTTG 3722
 5 TGGACTTTAT TAAGTAAAT CCCCTTCGCT GAAATTGCTT ATTTTTSCTG TTGGATAGAG 3782
 GATAGGGAGA ATATTTACTA ACTAAATACC ATTCAGTACT CATGCGTGAG ATGGGTGTAC 3842
 10 AAACATCATCC TCTTTTAATG GCATTTCTCT TTAAACTATG TTCCTAACCA AATGAGATGA 3902
 TAGGATAGAT CCTGGTTACC ACTCTTTTAC TGTGCACATA TGGGCCCCGG AATTC 3957

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1025 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

25 Met Ala Pro Val Leu Ser Lys Asp Ser Ala Asp Ile Glu Ser Ile Leu
 1 5 10 15
 30 Ala Leu Asn Pro Arg Thr Gln Thr His Ala Thr Leu Cys Ser Thr Ser
 20 25 30
 35 Ala Lys Lys Leu Asp Lys Lys His Trp Lys Arg Asn Pro Asp Lys Asn
 35 35 40 45
 Cys Phe Asn Cys Glu Lys Leu Glu Asn Asn Phe Asp Asp Ile Lys His
 50 55 60
 40 Thr Thr Leu Gly Glu Arg Gly Ala Leu Arg Glu Ala Met Arg Cys Leu
 65 70 75 80
 45 Lys Cys Ala Asp Ala Pro Cys Gln Lys Ser Cys Pro Thr Asn Leu Asp
 85 90 95
 Ile Lys Ser Phe Ile Thr Ser Ile Ala Asn Lys Asn Tyr Tyr Gly Ala
 100 105 110
 50 Ala Lys Met Ile Phe Ser Asp Asn Pro Leu Gly Leu Thr Cys Gly Met
 115 120 125
 Val Cys Pro Thr Ser Asp Leu Cys Val Gly Gly Cys Asn Leu Tyr Ala
 130 135 140
 55 Thr Glu Glu Gly Pro Ile Asn Ile Gly Gly Leu Gln Gln Phe Ala Thr
 145 150 155 160
 60 Glu Val Phe Lys Ala Met Ser Ile Pro Gln Ile Arg Asn Pro Ser Leu
 165 170 175
 Pro Pro Pro Glu Lys Met Ser Glu Ala Tyr Ser Ala Lys Ile Ala Leu
 180 185 190
 65 Phe Gly Ala Gly Pro Ala Ser Ile Ser Cys Ala Ser Phe Leu Ala Arg
 195 200 205
 Leu Gly Tyr Ser Asp Ile Thr Ile Phe Glu Lys Gln Glu Tyr Val Gly
 210 215 220
 Gly Leu Ser Thr Ser Glu Ile Pro Gln Phe Arg Leu Pro Tyr Asp Val
 225 230 235 240

50

	Asn	Ile	Glu	Leu	Ile	Ser	Glu	Lys	Thr	Ala	Ala	Tyr	Trp	Cys	Gln	Ser	
	610						615					620					
5	Val	Thr	Glu	Leu	Lys	Ala	Asp	Phe	Pro	Asp	Asn	Ile	Val	Ile	Ala	Ser	
	625					630					635					640	
	Ile	Met	Cys	Ser	Tyr	Asn	Lys	Asn	Asp	Trp	Thr	Glu	Leu	Ala	Lys	Lys	
					645					650					655		
10	Ser	Glu	Asp	Ser	Gly	Ala	Asp	Ala	Leu	Glu	Leu	Asn	Leu	Ser	Cys	Pro	
				660					665					670			
	His	Gly	Met	Gly	Glu	Arg	Gly	Met	Gly	Leu	Ala	Cys	Gly	Gln	Asp	Pro	
15			675					680					685				
	Glu	Leu	Val	Arg	Asn	Ile	Cys	Arg	Trp	Val	Arg	Gln	Ala	Val	Gln	Ile	
	690						695					700					
20	Pro	Phe	Phe	Ala	Lys	Leu	Thr	Pro	Asn	Val	Thr	Asp	Ile	Val	Ser	Ile	
	705					710					715					720	
	Ala	Arg	Ala	Ala	Lys	Glu	Gly	Gly	Ala	Asn	Gly	Val	Thr	Ala	Thr	Asn	
					725					730					735		
25	Thr	Val	Ser	Gly	Leu	Met	Gly	Leu	Lys	Ser	Asp	Gly	Thr	Pro	Trp	Pro	
				740					745					750			
	Ala	Val	Gly	Ile	Ala	Lys	Arg	Thr	Thr	Tyr	Gly	Gly	Val	Ser	Gly	Thr	
30			755					760					765				
	Ala	Ile	Arg	Pro	Ile	Ala	Leu	Arg	Ala	Val	Thr	Ser	Ile	Ala	Arg	Ala	
	770						775					780					
35	Leu	Pro	Gly	Phe	Pro	Ile	Leu	Ala	Thr	Gly	Gly	Ile	Asp	Ser	Ala	Glu	
	785					790					795					800	
	Ser	Gly	Leu	Gln	Phe	Leu	His	Ser	Gly	Ala	Ser	Val	Leu	Gln	Val	Cys	
					805					810					815		
40	Ser	Ala	Ile	Gln	Asn	Gln	Asp	Phe	Thr	Val	Ile	Glu	Asp	Tyr	Cys	Thr	
				820					825					830			
	Gly	Leu	Lys	Ala	Leu	Leu	Tyr	Leu	Lys	Ser	Ile	Glu	Glu	Leu	Gln	Asp	
45			835					840					845				
	Trp	Asp	Gly	Gln	Ser	Pro	Ala	Thr	Val	Ser	His	Gln	Lys	Gly	Lys	Pro	
	850						855					860					
50	Val	Pro	Arg	Ile	Ala	Glu	Leu	Met	Asp	Lys	Lys	Leu	Pro	Ser	Phe	Gly	
	865					870					875					880	
	Pro	Tyr	Leu	Glu	Gln	Arg	Lys	Lys	Ile	Ile	Ala	Glu	Asn	Lys	Ile	Arg	
					885					890					895		
55	Leu	Lys	Glu	Gln	Asn	Val	Ala	Phe	Ser	Pro	Leu	Lys	Arg	Ser	Cys	Phe	
				900					905					910			
	Ile	Pro	Lys	Arg	Pro	Ile	Pro	Thr	Ile	Lys	Asp	Val	Ile	Gly	Lys	Ala	
60			915					920					925				
	Leu	Gln	Tyr	Leu	Gly	Thr	Phe	Gly	Glu	Leu	Ser	Asn	Val	Glu	Gln	Val	
	930						935					940					
65	Val	Ala	Met	Ile	Asp	Glu	Glu	Met	Cys	Ile	Asn	Cys	Gly	Lys	Cys	Tyr	
	945					950					955					960	
	Met	Thr	Cys	Asn	Asp	Ser	Gly	Tyr	Gln	Ala	Ile	Gln	Phe	Asp	Pro	Glu	
					965					970					975		

51

Thr His Leu Pro Thr Ile Thr Asp Thr Cys Thr Gly Cys Thr Leu Cys
980 985 990

5 Leu Ser Val Cys Pro Ile Val Asp Cys Ile Lys Met Val Ser Arg Thr
995 1000 1005

Thr Pro Tyr Glu Pro Lys Arg Gly Val Pro Leu Ser Val Asn Pro Val
1010 1015 1020

10 Cys
1025

15

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4447 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 88..3162

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
 (B) LOCATION: 1..4447
 (D) OTHER INFORMATION: /product= "Pig DPD"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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25  GGACACTCGA CCCACGGCTC CCGCGGCGCG AGCGCGAGGA CGCGGGGAGG GCGCGCCGGT      60
    GGGAGACTCC AAGCTGTCTG CATCGCC ATG GCC CCT GTG CTG AGC AAG GAC      111
                               1      5
    GTG GCG GAC ATC GAG AGT ATC CTG GCT TTA AAT CCT CGA ACA CAG TCT      159
    Val Ala Asp Ile Glu Ser Ile Leu Ala Leu Asn Pro Arg Thr Gln Ser
        10      15      20
    CAT GCA GCC CTT CAT TCC ACT TTG GCC AAG AAA TTG GAT AAG AAA CAC      207
    His Ala Ala Leu His Ser Thr Leu Ala Lys Lys Leu Asp Lys Lys His
        25      30      35      40
    TGG AAA AGA AAT CCC GAT AAG AAC TGC TTT CAT TGC GAG AAG CTG GAG      255
    Trp Lys Arg Asn Pro Asp Lys Asn Cys Phe His Cys Glu Lys Leu Glu
        45      50      55
    AAT AAT TTT GGT GAC ATC AAG CAC ACG ACT CTT GGT GAG CGA GGA GCT      303
    Asn Asn Phe Gly Asp Ile Lys His Thr Leu Gly Glu Arg Gly Ala
        60      65      70
    CTC CGA GAA GCA ATG AGA TGC CTG AAA TGT GCC GAT GCT CCC TGT CAG      351
    Leu Arg Glu Ala Met Arg Cys Leu Lys Cys Ala Asp Ala Pro Cys Gln
        75      80      85
    AAG AGC TGT CCA ACT CAT CTA GAT ATC AAA TCA TTC ATC ACA AGT ATC      399
    Lys Ser Cys Pro Thr His Leu Asp Ile Lys Ser Phe Ile Thr Ser Ile
        90      95      100
    TCA AAT AAG AAC TAT TAT GGA GCT GCT AAG ATG ATT TTT TCT GAC AAC      447
    Ser Asn Lys Asn Tyr Tyr Gly Ala Ala Lys Met Ile Phe Ser Asp Asn
    105      110      115      120
    CCT CTT GGT CTG ACC TGT GGA ATG GTA TGT CCA ACC TCT GAT CTT TGT      495
    Pro Leu Gly Leu Thr Cys Gly Met Val Cys Pro Thr Ser Asp Leu Cys
        125      130      135
    GTA GGA GGA TGC AAT TTA TAT GCA ACT GAA GAG GGA TCA ATT AAT ATT      543
    Val Gly Gly Cys Asn Leu Tyr Ala Thr Glu Glu Gly Ser Ile Asn Ile
        140      145      150
    GGT GGA TTG CAG CAG TTT GCT TCT GAG GTG TTC AAA GCA ATG AAT ATC      591
    Gly Gly Leu Gln Gln Phe Ala Ser Glu Val Phe Lys Ala Met Asn Ile
        155      160      165

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	CCA	CAA	ATC	AGG	AAT	CCT	TGT	CTG	CCA	TCC	CAA	GAG	AAA	ATG	CCT	GAA	639
	Pro	Gln	Ile	Arg	Asn	Pro	Cys	Leu	Pro	Ser	Gln	Glu	Lys	Met	Pro	Glu	
	170						175					180					
5	GCT	TAT	TCT	GCA	AAG	ATT	GCT	CTT	TTG	GGT	GCT	GGG	CCT	GCA	AGT	ATA	687
	Ala	Tyr	Ser	Ala	Lys	Ile	Ala	Leu	Leu	Gly	Ala	Gly	Pro	Ala	Ser	Ile	
	185					190					195					200	
10	AGC	TGT	GCT	TCC	TTG	TTG	GCT	CGA	TTA	GGC	TAC	TCT	GAC	ATC	ACT	ATA	735
	Ser	Cys	Ala	Ser	Phe	Leu	Ala	Arg	Leu	Gly	Tyr	Ser	Asp	Ile	Thr	Ile	
					205					210					215		
15	TTT	GAA	AAA	CAA	GAA	TAT	GTT	GGT	GGT	TTA	AGT	ACT	TCT	GAA	ATC	CCT	783
	Phe	Glu	Lys	Gln	Glu	Tyr	Val	Gly	Gly	Leu	Ser	Thr	Ser	Glu	Ile	Pro	
				220				225						230			
20	CAG	TTC	CGG	CTG	CCA	TAT	GAT	GTA	GTG	AAT	TTT	GAG	ATT	GAG	CTT	ATG	831
	Gln	Phe	Arg	Leu	Pro	Tyr	Asp	Val	Val	Asn	Phe	Glu	Ile	Glu	Leu	Met	
			235					240					245				
	AAG	GAC	CTT	GGT	GTA	AAG	ATA	ATT	TGT	GGT	AAA	AGC	CTT	TCA	GAG	AAT	879
	Lys	Asp	Leu	Gly	Val	Lys	Ile	Ile	Cys	Gly	Lys	Ser	Leu	Ser	Glu	Asn	
		250					255					260					
25	GAA	ATT	ACT	CTC	AAG	ACT	TTA	AAA	GAA	GAA	GGG	TAT	AAA	GCT	GCT	TTC	927
	Glu	Ile	Thr	Leu	Asn	Thr	Leu	Lys	Glu	Glu	Gly	Tyr	Lys	Ala	Ala	Phe	
	265					270					275					280	
30	ATT	GGT	ATA	GGT	TTG	CCA	GAA	CCC	AAA	ACG	GAT	GAC	ATC	TTC	CAA	GGC	975
	Ile	Gly	Ile	Gly	Leu	Pro	Glu	Pro	Lys	Thr	Asp	Asp	Ile	Phe	Gln	Gly	
					285					290					295		
35	CTG	ACA	CAG	GAC	CAG	GGG	TTT	TAC	ACA	TCC	AAA	GAC	TTT	CTG	CCC	CTT	1023
	Leu	Thr	Gln	Asp	Gln	Gly	Phe	Tyr	Thr	Ser	Lys	Asp	Phe	Leu	Pro	Leu	
				300					305					310			
40	GTA	GCC	AAA	AGC	AGT	AAA	GCA	GGA	ATG	TGT	GCC	TGT	CAC	TCT	CCA	TTG	1071
	Val	Ala	Lys	Ser	Ser	Lys	Ala	Gly	Met	Cys	Ala	Cys	His	Ser	Pro	Leu	
			315					320					325				
	CCA	TCG	ATA	CGG	GGA	GCC	GTG	ATT	GTA	CTC	GGA	GCT	GGA	GAC	ACA	GCT	1119
	Pro	Ser	Ile	Arg	Gly	Ala	Val	Ile	Val	Leu	Gly	Ala	Gly	Asp	Thr	Ala	
			330				335					340					
45	TTC	GAC	TGT	GCA	ACA	TCC	GCT	TTA	CGT	TGT	GGA	GCC	CGC	CGA	GTG	TTC	1167
	Phe	Asp	Cys	Ala	Thr	Ser	Ala	Leu	Arg	Cys	Gly	Ala	Arg	Arg	Val	Phe	
	345					350					355					360	
50	CTC	GTC	TTC	AGA	AAA	GGC	TTT	GTT	AAT	ATA	AGA	GCT	GTC	CCT	GAG	GAG	1215
	Leu	Val	Phe	Arg	Lys	Gly	Phe	Val	Asn	Ile	Arg	Ala	Val	Pro	Glu	Glu	
					365					370					375		
55	GTG	GAG	CTT	GCT	AAG	GAA	GAA	AAA	TGT	GAA	TTT	TTG	CCT	TTC	CTG	TCC	1263
	Val	Glu	Leu	Ala	Lys	Glu	Glu	Lys	Cys	Glu	Phe	Leu	Pro	Phe	Leu	Ser	
				380					385					390			
	CCA	CGG	AAG	GTT	ATA	GTT	AAA	GGT	GGG	AGA	ATT	GTT	GCC	GTG	CAA	TTT	1311
	Pro	Arg	Lys	Val	Ile	Val	Lys	Gly	Gly	Arg	Ile	Val	Ala	Val	Gln	Phe	
			395					400					405				
60	GTT	CGA	ACA	GAA	CAA	GAT	GAA	ACT	GGA	AAA	TGG	AAT	GAA	GAT	GAA	GAT	1359
	Val	Arg	Thr	Glu	Gln	Asp	Glu	Thr	Gly	Lys	Trp	Asn	Glu	Asp	Glu	Asp	
			410				415					420					
65	CAG	ATA	GTC	CAT	CTG	AAG	GCT	GAT	GTG	GTC	ATC	AGT	GCC	TTT	GGC	TCA	1407
	Gln	Ile	Val	His	Leu	Lys	Ala	Asp	Val	Val	Ile	Ser	Ala	Phe	Gly	Ser	
	425					430					435					440	
	GTG	CTG	AGG	GAT	CCT	AAA	GTA	AAA	GAA	GCC	TTG	AGC	CCT	ATA	AAA	TTT	1455

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	Val	Leu	Arg	Asp	Pro	Lys	Val	Lys	Glu	Ala	Leu	Ser	Pro	Ile	Lys	Phe	
				445						450					455		
5	AAC	AGA	TGG	GAT	CTC	CCA	GAA	GTA	GAT	CCA	GAA	ACT	ATG	CAA	ACC	AGT	1503
	Asn	Arg	Trp	Asp	Leu	Pro	Glu	Val	Asp	Pro	Glu	Thr	Met	Gln	Thr	Ser	
				460					465					470			
10	GAA	CCA	TGG	GTG	TTT	GCA	GGT	GGT	GAT	ATC	GTT	GGT	ATG	GCT	AAC	ACT	1551
	Glu	Pro	Trp	Val	Phe	Ala	Gly	Gly	Asp	Ile	Val	Gly	Met	Ala	Asn	Thr	
				475				480					485				
15	ACG	GTG	GAA	TCC	GTA	AAT	GAC	GGA	AAG	CAG	GCC	TCC	TGG	TAC	ATT	CAC	1599
	Thr	Val	Glu	Ser	Val	Asn	Asp	Gly	Lys	Gln	Ala	Ser	Trp	Tyr	Ile	His	
				490			495					500					
20	AAA	TAT	ATC	CAG	GCC	CAA	TAT	GGA	GCT	TCA	GTT	TCT	GCC	AAG	CCC	GAA	1647
	Lys	Tyr	Ile	Gln	Ala	Gln	Tyr	Gly	Ala	Ser	Val	Ser	Ala	Lys	Pro	Glu	
	505					510					515					520	
25	CTG	CCC	CTG	TTT	TAT	ACG	CCT	GTT	GAC	CTG	GTG	GAC	ATC	AGC	GTG	GAA	1695
	Leu	Pro	Leu	Phe	Tyr	Thr	Pro	Val	Asp	Leu	Val	Asp	Ile	Ser	Val	Glu	
					525					530					535		
30	ATG	GCT	GGA	TTA	AAG	TTT	ATA	AAT	CCT	TTT	GGT	CTT	GCC	AGT	GCA	GCT	1743
	Met	Ala	Gly	Leu	Lys	Phe	Ile	Asn	Pro	Phe	Gly	Leu	Ala	Ser	Ala	Ala	
				540					545					550			
35	CCA	ACT	ACC	AGT	TCA	TCG	ATG	ATT	CGA	AGA	GCT	TTT	GAA	GCT	GGA	TGG	1791
	Pro	Thr	Ser	Ser	Ser	Ser	Met	Ile	Arg	Arg	Ala	Phe	Glu	Ala	Gly	Trp	
				555				560					565				
40	GGT	TTT	GCC	CTG	ACC	AAA	ACT	TTC	TCT	CTT	GAT	AAG	GAC	ATA	GTG	ACA	1839
	Gly	Phe	Ala	Leu	Thr	Lys	Thr	Phe	Ser	Leu	Asp	Lys	Asp	Ile	Val	Thr	
				570			575					580					
45	AAT	GTC	TCA	CCC	AGA	ATC	GTC	CGG	GGG	ACT	ACC	TCT	GGC	CCC	ATG	TAC	1887
	Asn	Val	Ser	Pro	Arg	Ile	Val	Arg	Gly	Thr	Thr	Ser	Gly	Pro	Met	Tyr	
	585					590					595					600	
50	GGC	CCT	GGA	CAA	AGC	TCC	TTC	CTG	AAT	ATT	GAG	CTC	ATC	AGT	GAA	AAA	1935
	Gly	Pro	Gly	Gln	Ser	Ser	Phe	Leu	Asn	Ile	Glu	Leu	Ile	Ser	Glu	Lys	
					605					610					615		
55	ACA	GCT	GCA	TAT	TGG	TGT	CAA	AGT	GTC	ACT	GAA	CTA	AAA	GCT	GAC	TTT	1983
	Thr	Ala	Ala	Tyr	Trp	Cys	Gln	Ser	Val	Thr	Glu	Leu	Lys	Ala	Asp	Phe	
				620					625					630			
60	CCA	GAC	AAT	ATT	GTG	ATC	GCC	AGC	ATC	ATG	TGT	AGT	TAC	AAC	AAA	AAT	2031
	Pro	Asp	Asn	Ile	Val	Ile	Ala	Ser	Ile	Met	Cys	Ser	Tyr	Asn	Lys	Asn	
				635				640					645				
65	GAC	TGG	ATG	GAA	CTC	TCC	AGA	AAG	GCT	GAG	GCC	TCT	GGA	GCA	GAT	GCC	2079
	Asp	Trp	Met	Glu	Leu	Ser	Arg	Lys	Ala	Glu	Ala		Gly	Ala	Asp	Ala	
				650			655					660					
70	TTG	GAG	TTA	AAT	CTG	TCA	TGT	CCA	CAC	GGC	ATG	GGA	GAA	AGA	GGA	ATG	2127
	Leu	Glu	Leu	Asn	Leu	Ser	Cys	Pro	His	Gly	Met	Gly	Glu	Arg	Gly	Met	
						670					675					680	
75	GGC	CTG	GCT	TGT	GGG	CAG	GAT	CCA	GAG	CTG	GTG	CGG	AAC	ATC	TGT	CGC	2175
	Gly	Leu	Ala	Cys	Gly	Gln	Asp	Pro	Glu	Leu	Val	Arg	Asn	Ile	Cys	Arg	
					685					690					695		
80	TGG	GTT	AGG	CAA	GCT	GTT	CAG	ATT	CCC	TTT	TTT	GCC	AAG	TTG	ACC	CCA	2223
	Trp	Val	Arg	Gln	Ala	Val	Gln	Ile	Pro	Phe	Phe	Ala	Lys	Leu	Thr	Pro	
				700					705					710			
85	AAC	GTC	ACT	GAT	ATA	GTA	AGC	ATC	GCC	AGA	GCG	GCC	AAG	GAA	GGT	GGC	2271

55

	Asn	Val	Thr	Asp	Ile	Val	Ser	Ile	Ala	Arg	Ala	Ala	Lys	Glu	Gly	Gly	
		715						720					725				
5	GCA	GAT	GGT	GTT	ACA	GCC	ACC	AAC	ACG	GTC	TCA	GGT	CTC	ATG	GGA	TTA	2319
	Ala	Asp	Gly	Val	Thr	Ala	Thr	Asn	Thr	Val	Ser	Gly	Leu	Met	Gly	Leu	
		730					735					740					
10	AAA	GCC	GAT	GGC	ACG	CCC	TGG	CCA	GCG	GTG	GGT	GCT	GGC	AAG	CGG	ACT	2367
	Lys	Ala	Asp	Gly	Thr	Pro	Trp	Pro	Ala	Val	Gly	Ala	Gly	Lys	Arg	Thr	
		745				750				755						760	
15	ACA	TAC	GGA	GGA	GTG	TCT	GGC	ACG	GCC	ATC	AGA	CCA	ATT	GCT	TTG	AGA	2415
	Thr	Tyr	Gly	Gly	Val	Ser	Gly	Thr	Ala	Ile	Arg	Pro	Ile	Ala	Leu	Arg	
					765					770					775		
20	GCT	GTG	ACC	ACC	ATT	GCT	CGT	GCT	TTG	CCT	GGA	TTT	CCC	ATT	TTG	GCT	2463
	Ala	Val	Thr	Thr	Ile	Ala	Arg	Ala	Leu	Pro	Gly	Phe	Pro	Ile	Leu	Ala	
					780				785					790			
25	ACT	GGT	GGA	ATT	GAC	TCA	GCT	GAA	AGT	GGA	CTT	CAG	TTT	CTC	CAC	AGT	2511
	Thr	Gly	Gly	Ile	Asp	Ser	Ala	Glu	Ser	Gly	Leu	Gln	Phe	Leu	His	Ser	
			795					800					805				
30	GGT	GCT	TCG	GTC	CTC	CAG	GTA	TGC	AGT	GCT	GTT	CAG	AAT	CAG	GAT	TTC	2559
	Gly	Ala	Ser	Val	Leu	Gln	Val	Cys	Ser	Ala	Val	Gln	Asn	Gln	Asp	Phe	
		810					815					820					
35	ACT	GTC	ATC	CAA	GAC	TAT	TGC	ACT	GGC	CTC	AAA	GCC	TTG	CTT	TAT	CTG	2607
	Thr	Val	Ile	Gln	Asp	Tyr	Cys	Thr	Gly	Leu	Lys	Ala	Leu	Leu	Tyr	Leu	
		825				830					835					840	
40	AAA	AGC	ATT	GAA	GAA	CTA	CAA	GGC	TGG	GAT	GGG	CAG	AGT	CCA	GGT	ACC	2655
	Lys	Ser	Ile	Glu	Glu	Leu	Gln	Gly	Trp	Asp	Gly	Gln	Ser	Pro	Gly	Thr	
					845					850					855		
45	GAG	AGT	CAC	CAG	AAG	GGG	AAA	CCA	GTT	CCT	CGT	ATT	GCT	GAA	CTC	ATG	2703
	Glu	Ser	His	Gln	Lys	Gly	Lys	Pro	Val	Pro	Arg	Ile	Ala	Glu	Leu	Met	
					860				865					870			
50	GGA	AAG	AAA	CTG	CCA	AAT	TTT	GGA	CCT	TAT	CTG	GAG	CAA	CGC	AAG	AAA	2751
	Gly	Lys	Lys	Leu	Pro	Asn	Phe	Gly	Pro	Tyr	Leu	Glu	Gln	Arg	Lys	Lys	
			875					880					885				
55	ATC	ATA	GCA	GAG	GAA	AAG	ATG	AGA	CTG	AAA	GAA	CAA	AAT	GCA	GCT	TTT	2799
	Ile	Ile	Ala	Glu	Glu	Lys	Met	Arg	Leu	Lys	Glu	Gln	Asn	Ala	Ala	Phe	
		890					895					900					
60	CCA	CCA	CTT	GAG	AGA	AAA	CCT	TTT	ATT	CCC	AAA	AAG	CCT	ATT	CCT	GCT	2847
	Pro	Pro	Leu	Glu	Arg	Lys	Pro	Phe	Ile	Pro	Lys	Lys	Pro	Ile	Pro	Ala	
		905				910					915					920	
65	ATT	AAG	GAT	GTA	ATT	GGA	AAA	GCA	CTG	CAG	TAC	CTT	GGA	ACG	TTT	GGT	2895
	Ile	Lys	Asp	Val	Ile	Gly	Lys	Ala	Leu	Gln	Tyr	Leu	Gly	Thr	Phe	Gly	
					925					930					935		
70	GAA	CTG	AGC	AAC	ATA	GAG	CAA	GTT	GTG	GCT	GTG	ATC	GAT	GAA	GAA	ATG	2943
	Glu	Leu	Ser	Asn	Ile	Glu	Gln	Val	Val	Ala	Val	Ile	Asp	Glu	Glu	Met	
					940				945					950			
75	TGT	ATC	AAC	TGT	GGC	AAA	TGC	TAC	ATG	ACC	TGT	AAT	GAC	TCT	GGC	TAC	2991
	Cys	Ile	Asn	Cys	Gly	Lys	Cys	Tyr	Met	Thr	Cys	Asn	Asp	Ser	Gly	Tyr	
			955					960					965				
80	CAG	GCT	ATC	CAG	TTT	GAT	CCC	GAA	ACC	CAC	CTG	CCC	ACC	GTT	ACT	GAC	3039
	Gln	Ala	Ile	Gln	Phe	Asp	Pro	Glu	Thr	His	Leu	Pro	Thr	Val	Thr	Asp	
		970					975					980					
85	ACT	TGC	ACA	GGC	TGT	ACC	CTG	TGT	CTC	TCC	GTC	TGC	CCT	ATT	ATC	GAC	3087

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	Thr Cys Thr Gly Cys Thr Leu Cys Leu Ser Val Cys Pro Ile Ile Asp	
	985 990 995 1000	
5	TGC ATC AGA ATG GTT TCC AGG ACA ACA CCT TAC GAA CCA AAG AGA GGC Cys Ile Arg Met Val Ser Arg Thr Thr Pro Tyr Glu Pro Lys Arg Gly	3135
	1005 1010 1015	
10	TTG CCC TTG GCT GTG AAT CCG GTG TGC TGAGGTGATT CGTGGAAACAG Leu Pro Leu Ala Val Asn Pro Val Cys	3182
	1020 1025	
	TTGCTGTGAA CTTTGAGGTC ACCCCCATAT GCTGTCTTTT TAATTGTGGT TATTATACTC	3242
15	AGCTCTTTCT CAATGAAAAC AAATATAATA TTTCTAGATA AAAGTTCTAA ATACATGTCT	3302
	AAATTTTAAA AAACATCTAC TGCCAGAGCC CGTTCAATTA ATGGTCATAA AATAGAATCC	3362
	TGCTTTTCTG AGGCTAGTTG TTCAATAACT GCTGCAGTTA ATTGGATGTT CTCCATCAGT	3422
20	TATCCATTAT GAAAAATATT AACTTTTTTG GTGGCAATTT CCAAATTGCC CTATGCTGTG	3482
	CTCTGTCTTT GATTTCTAAT TGTAAGTGAA GTTAAGCATT TTAGAACAAA GTATAATTTA	3542
25	ACTTTCAAGC AAATGTTTCC AAGGAAACAT TTTATAATTA AAAATTACAA TTTAATTTTA	3602
	ACACTGTTCC TAAGCAAATG TAATTAGCTC CATAAAGCTC AAATGAAGTC AAATAATTAT	3662
	TTACTGTGGC AGGAAAAGAA AGCCAATGAG GGTTCGCAAA ACTTCTCTAA GGCCCTTTGG	3722
30	CTGAAATAAC TTCTCTTTGG TGCTACATAC TGAAAGTGAC TGTTTAATCA TCATTCATGT	3782
	CACACCGTGC TCCCTCGCCC TCAGGCCTGA GATGGGTCTC CAGACTCCAC CAGTGAATCA	3842
35	GCATGACACC TTCTTTAACT GTGTGAGCGA CGTTCCTAAC AAAGTAAGGT GTGGGGATGA	3902
	AGCTCTGGTT AAAGCCACTC TTTTGCTGTG CTCCGATCTG TTCTATCCGC TTCTGAGAGC	3962
	AACCTTCATG ATTACAGCAA TTAATGTTTG CACAGAGCCC AGATTATACA GCAGTGGGTC	4022
40	ATTGTGCTTC ATTATTCAAG AATGAAGATA AAGACAAATA GAGGATTAGT AAAATATATT	4082
	AAATGTGCAA TACCACTTAA ATGACTCTTA ATGTTTATAT TGAATTTCCA AAGCGATTAA	4142
45	ATAAAAAAGA GCTATTTTTT GTTATTGCCA AACAATATTT TTTGTATTTT TCTATTTTCA	4202
	TAATGAGCAA ATAGCATCCT ATAAATCTGT TTATCTCTTC TTTGTAGTGT GTTTTCATAT	4262
	AAATCCACAA GTAGAAAATC TTTTCATCTG TGGCATATTT CTATGACAAA TGCAAGATCT	4322
50	AGAAAAATTA AATGTTTGAT TATGCCATTT TGGAATGCA TATTTACCAC CAAACCTATG	4382
	TGACTGAATA ATGTCAAATA AAATTTTATG AATCATTTTA AAAAAAAAAA AAAAAGGGCG	4442
55	GCCGC	4447
60		

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3025 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ala Pro Val Leu Ser Lys Asp Val Ala Asp Ile Glu Ser Ile Leu
 1 5 10 15
 Ala Leu Asn Pro Arg Thr Gln Ser His Ala Ala Leu His Ser Thr Leu
 20 25 30
 Ala Lys Lys Leu Asp Lys Lys His Trp Lys Arg Asn Pro Asp Lys Asn
 35 40 45
 Cys Phe His Cys Glu Lys Leu Glu Asn Asn Phe Gly Asp Ile Lys His
 50 55 60
 Thr Thr Leu Gly Glu Arg Gly Ala Leu Arg Glu Ala Met Arg Cys Leu
 65 70 75 80
 Lys Cys Ala Asp Ala Pro Cys Gln Lys Ser Cys Pro Thr His Leu Asp
 85 90 95
 Ile Lys Ser Phe Ile Thr Ser Ile Ser Asn Lys Asn Tyr Tyr Gly Ala
 100 105 110
 Ala Lys Met Ile Phe Ser Asp Asn Pro Leu Gly Leu Thr Cys Gly Met
 115 120 125
 Val Cys Pro Thr Ser Asp Leu Cys Val Gly Gly Cys Asn Leu Tyr Ala
 130 135 140
 Thr Glu Glu Gly Ser Ile Asn Ile Gly Gly Leu Gln Gln Phe Ala Ser
 145 150 155 160
 Glu Val Phe Lys Ala Met Asn Ile Pro Gln Ile Arg Asn Pro Cys Leu
 165 170 175
 Pro Ser Gln Glu Lys Met Pro Glu Ala Tyr Ser Ala Lys Ile Ala Leu
 180 185 190
 Leu Gly Ala Gly Pro Ala Ser Ile Ser Cys Ala Ser Phe Leu Ala Arg
 195 200 205
 Leu Gly Tyr Ser Asp Ile Thr Ile Phe Glu Lys Gln Glu Tyr Val Gly
 210 215 220
 Gly Leu Ser Thr Ser Glu Ile Pro Gln Phe Arg Leu Pro Tyr Asp Val
 225 230 235 240
 Val Asn Phe Glu Ile Glu Leu Met Lys Asp Leu Gly Val Lys Ile Ile
 245 250 255
 Cys Gly Lys Ser Leu Ser Glu Asn Glu Ile Thr Leu Asn Thr Leu Lys
 260 265 270
 Glu Glu Gly Tyr Lys Ala Ala Phe Ile Gly Ile Gly Leu Pro Glu Pro
 275 280 285
 Lys Thr Asp Asp Ile Phe Gln Gly Leu Thr Gln Asp Gln Gly Phe Tyr
 290 295 300

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	Thr	Ser	Lys	Asp	Phe	Leu	Pro	Leu	Val	Ala	Lys	Ser	Ser	Lys	Ala	Gly
	305					310					315					320
5	Met	Cys	Ala	Cys	His	Ser	Pro	Leu	Pro	Ser	Ile	Arg	Gly	Ala	Val	Ile
					325					330					335	
	Val	Leu	Gly	Ala	Gly	Asp	Thr	Ala	Phe	Asp	Cys	Ala	Thr	Ser	Ala	Leu
				340					345					350		
10	Arg	Cys	Gly	Ala	Arg	Arg	Val	Phe	Leu	Val	Phe	Arg	Lys	Gly	Phe	Val
			355					360					365			
	Asn	Ile	Arg	Ala	Val	Pro	Glu	Glu	Val	Glu	Leu	Ala	Lys	Glu	Glu	Lys
15		370					375					380				
	Cys	Glu	Phe	Leu	Pro	Phe	Leu	Ser	Pro	Arg	Lys	Val	Ile	Val	Lys	Gly
	385					390					395					400
20	Gly	Arg	Ile	Val	Ala	Val	Gln	Phe	Val	Arg	Thr	Glu	Gln	Asp	Glu	Thr
					405					410					415	
	Gly	Lys	Trp	Asn	Glu	Asp	Glu	Asp	Gln	Ile	Val	His	Leu	Lys	Ala	Asp
				420					425					430		
25	Val	Val	Ile	Ser	Ala	Phe	Gly	Ser	Val	Leu	Arg	Asp	Pro	Lys	Val	Lys
			435					440					445			
	Glu	Ala	Leu	Ser	Pro	Ile	Lys	Phe	Asn	Arg	Trp	Asp	Leu	Pro	Glu	Val
30		450					455					460				
	Asp	Pro	Glu	Thr	Met	Gln	Thr	Ser	Glu	Pro	Trp	Val	Phe	Ala	Gly	Gly
	465					470					475					480
35	Asp	Ile	Val	Gly	Met	Ala	Asn	Thr	Thr	Val	Glu	Ser	Val	Asn	Asp	Gly
					485					490					495	
	Lys	Gln	Ala	Ser	Trp	Tyr	Ile	His	Lys	Tyr	Ile	Gln	Ala	Gln	Tyr	Gly
				500					505					510		
40	Ala	Ser	Val	Ser	Ala	Lys	Pro	Glu	Leu	Pro	Leu	Phe	Tyr	Thr	Pro	Val
			515					520					525			
	Asp	Leu	Val	Asp	Ile	Ser	Val	Glu	Met	Ala	Gly	Leu	Lys	Phe	Ile	Asn
45		530					535					540				
	Pro	Phe	Gly	Leu	Ala	Ser	Ala	Ala	Pro	Thr	Thr	Ser	Ser	Ser	Met	Ile
	545					550					555					560
50	Arg	Arg	Ala	Phe	Glu	Ala	Gly	Trp	Gly	Phe	Ala	Leu	Thr	Lys	Thr	Phe
					565					570					575	
	Ser	Leu	Asp	Lys	Asp	Ile	Val	Thr	Asn	Val	Ser	Pro	Arg	Ile	Val	Arg
				580					585					590		
55	Gly	Thr	Thr	Ser	Gly	Pro	Met	Tyr	Gly	Pro	Gly	Gln	Ser	Ser	Phe	Leu
			595					600					605			
	Asn	Ile	Glu	Leu	Ile	Ser	Glu	Lys	Thr	Ala	Ala	Tyr	Trp	Cys	Gln	Ser
60		610					615					620				
	Val	Thr	Glu	Leu	Lys	Ala	Asp	Phe	Pro	Asp	Asn	Ile	Val	Ile	Ala	Ser
	625					630					635					640
65	Ile	Met	Cys	Ser	Tyr	Asn	Lys	Asn	Asp	Trp	Met	Glu	Leu	Ser	Arg	Lys
					645					650					655	
	Ala	Glu	Ala	Ser	Gly	Ala	Asp	Ala	Leu	Glu	Leu	Asn	Leu	Ser	Cys	Pro
					660				665					670		

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His Gly Met Gly Glu Arg Gly Met Gly Leu Ala Cys Gly Gln Asp Pro
 575 580 585
 5 Glu Leu Val Arg Asn Ile Cys Arg Trp Val Arg Gln Ala Val Gln Ile
 690 695 700
 Pro Phe Phe Ala Lys Leu Thr Pro Asn Val Thr Asp Ile Val Ser Ile
 705 710 715 720
 10 Ala Arg Ala Ala Lys Glu Gly Gly Ala Asp Gly Val Thr Ala Thr Asn
 725 730 735
 15 Thr Val Ser Gly Leu Met Gly Leu Lys Ala Asp Gly Thr Pro Trp Pro
 740 745 750
 Ala Val Gly Ala Gly Lys Arg Thr Thr Tyr Gly Gly Val Ser Gly Thr
 755 760 765
 20 Ala Ile Arg Pro Ile Ala Leu Arg Ala Val Thr Thr Ile Ala Arg Ala
 770 775 780
 Leu Pro Gly Phe Pro Ile Leu Ala Thr Gly Gly Ile Asp Ser Ala Glu
 785 790 795 800
 25 Ser Gly Leu Gln Phe Leu His Ser Gly Ala Ser Val Leu Gln Val Cys
 805 810 815
 30 Ser Ala Val Gln Asn Gln Asp Phe Thr Val Ile Gln Asp Tyr Cys Thr
 820 825 830
 Gly Leu Lys Ala Leu Leu Tyr Leu Lys Ser Ile Glu Glu Leu Gln Gly
 835 840 845
 35 Trp Asp Gly Gln Ser Pro Gly Thr Glu Ser His Gln Lys Gly Lys Pro
 850 855 860
 Val Pro Arg Ile Ala Glu Leu Met Gly Lys Lys Leu Pro Asn Phe Gly
 865 870 875 880
 40 Pro Tyr Leu Glu Gln Arg Lys Lys Ile Ile Ala Glu Glu Lys Met Arg
 885 890 895
 45 Leu Lys Glu Gln Asn Ala Ala Phe Pro Pro Leu Glu Arg Lys Pro Phe
 900 905 910
 Ile Pro Lys Lys Pro Ile Pro Ala Ile Lys Asp Val Ile Gly Lys Ala
 915 920 925
 50 Leu Gln Tyr Leu Gly Thr Phe Gly Glu Leu Ser Asn Ile Glu Gln Val
 930 935 940
 Val Ala Val Ile Asp Glu Glu Met Cys Ile Asn Cys Gly Lys Cys Tyr
 945 950 955 960
 55 Met Thr Cys Asn Asp Ser Gly Tyr Gln Ala Ile Gln Phe Asp Pro Glu
 965 970 975
 60 Thr His Leu Pro Thr Val Thr Asp Thr Cys Thr Gly Cys Thr Leu Cys
 980 985 990
 Leu Ser Val Cys Pro Ile Ile Asp Cys Ile Arg Met Val Ser Arg Thr
 995 1000 1005
 65 Thr Pro Tyr Glu Pro Lys Arg Gly Leu Pro L u Ala Val Asn Pro Val
 1010 1015 1020
 Cys
 1025

- 5 (2) INFORMATION FOR SEQ ID NO:5:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
10 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (primer)
- 15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
GCAAGGAGGG TTTGTCACTG 20
- 20 (2) INFORMATION FOR SEQ ID NO:6:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 base pairs
 (B) TYPE: nucleic acid
25 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (primer)
- 30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
CCGATTCCAC TGTAGTGTTA GCC 23
- 35 (2) INFORMATION FOR SEQ ID NO:7:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
40 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (primer)
- 45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
TAACACTACA GTGGAATCGG 20
- 50 (2) INFORMATION FOR SEQ ID NO:8:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
55 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (primer)
- 60 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
AAATCCAGGC AGAGCACGAG 20
- 65 (2) INFORMATION FOR SEQ ID NO:9:

(ii) MOLECULE TYPE: DNA (primer).

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

24

(2) INFORMATION FOR SEO ID NO:10:

(11) MOLECULE TYPE: DNA (primer)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

24

(2) INFORMATION FOR SEQ ID NO:11:

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

(2) INFORMATION FOR SEQ ID NO:12:

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 17 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single

62

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

10

Val Xaa Val Xaa Gly Xaa Gly Xaa Xaa Gly Xaa Xaa Xaa Ala Xaa Xaa
1 5 10 15
Ala

WHAT IS CLAIMED IS:

1 1. An isolated nucleic acid encoding a dihydropyrimidine
2 dehydrogenase (DPD) protein, said nucleic acid capable of selectively hybridizing to
3 a second nucleic acid consisting of the nucleotide sequence of Seq. ID. No. 1 or
4 Seq. ID No. 3 under stringent hybridization conditions.

1 2. The nucleic acid of claim 1 wherein the nucleic acid is of
2 human origin.

1 3. The nucleic acid of claim 2 wherein the nucleic acid consists of
2 the nucleotide sequence of Seq. ID. No. 1.

1 4. The nucleic acid of claim 1 wherein the nucleic acid is of pig
2 origin.

1 5. The nucleic acid of claim 4 wherein the nucleic acid consists of
2 the nucleotide sequence of Seq. ID. No. 3.

1 6. The nucleic acid of claim 1 wherein the nucleic acid is full-
2 length.

1 7. An isolated nucleic acid that codes for a DPD polypeptide,
2 wherein a polypeptide expressed from the nucleic acid specifically binds to an
3 antibody generated against an immunogen consisting of a DPD polypeptide having
4 an amino acid sequence as depicted by Seq. ID No. 2 or Seq. ID No. 4.

1 8. The nucleic acid of claim 7 wherein the nucleic acid is of
2 human origin.

1 9. The nucleic acid of claim 8 wherein said nucleic acid consists
2 of the polynucleotide sequence of Seq. ID. No. 1.

1 10. The nucleic acid of claim 7 wher in said nucleic acid is of pig
2 origin.

1 11. The nucleic acid of claim 10 wherein said nucleic acid consists
2 of the polynucleotide sequence of Seq. ID No. 3.

1 12. The nucleic acid of claim 7 wherein said nucleic acid is full-
2 length.

1 13. An oligonucleotide probe that is capable of selectively
2 hybridizing, under stringent hybridizing conditions, to a DPD nucleic acid having a
3 nucleotide sequence of Seq. ID No. 1 or Seq. ID No. 3.

1 14. An oligonucleotide probe of claim 13 that is between about 10
2 and 100 nucleotides in length.

1 15. A method for determining whether a patient is at risk of a
2 toxic reaction to 5-fluorouracil, the method comprising analyzing DPD DNA or
3 mRNA in a sample from the patient to determine the amount of intact DPD nucleic
4 acid, wherein an enhanced risk of a toxic reaction to 5-fluorouracil is indicated by a
5 decrease in the amount of intact DPD DNA or mRNA in the sample compared to the
6 amount of DPD DNA or mRNA in a sample obtained from a patient known to not
7 have a DPD deficiency.

1 16. A method of claim 15 wherein an enhanced risk of a toxic
2 reaction is indicated by a decrease of greater than about 70%.

1 17. A method of claim 15 wherein an increased risk of a toxic
2 reaction is indicated by a decrease of greater than about 50%.

1 18. The method of claim 15, wherein the method comprises the
2 steps of:

- 3 (a) obtaining a cellular sample from the patient;
4 (b) extracting DNA or RNA from the sample;
5 (c) hybridizing a probe comprising a DPD nucleic acid to the
6 DNA or RNA from the sample; and

(d) determining whether the DPD nucleic acid binds to the DNA or RNA.

19. The method of claim 15, wherein the DPD nucleic acid is analyzed by RT-PCR.

20. The method of claim 15, wherein the DPD nucleic acid is analyzed by PCR sequencing of genomic DNA from the patient.

21. A method of claim 15 wherein the cellular sample comprises lymphocytes.

22. A method of claim 15 wherein the probe oligonucleotide probe that is capable of selectively hybridizing, under stringent hybridizing conditions, to a DPD nucleic acid having a nucleotide sequence or a specific subsequence of that shown in Seq. ID No. 1 or Seq. ID No. 3.

23. A method of claim 22 wherein the oligonucleotide probe is between about 10 and 100 nucleotides in length.

24. A method for expressing recombinant DPD protein in a prokaryotic cell, the method comprising the steps of:

a) transfecting the cell with an expression vector comprising a promoter that is operably linked to a nucleic acid that encodes DPD; and

b) incubating the cell in a medium that contains uracil to allow expression of the recombinant DPD protein.

25. A method of claim 24 wherein the medium contains about 100 μ M uracil.

26. A method of claim 24 wherein the medium contains 100 μ M each of FAD and FMN.

1 27. An expression vector comprising a selectable marker, wherein
2 the selectable marker is a nucleic acid that encodes DPD.

1 28. An expression vector as in claim 27 wherein the selectable
2 marker is operably linked to at least one promoter.

1 29. An expression vector as in claim 28 wherein the promoter
2 functions in a eukaryote.

1 30. An expression vector as in claim 28 wherein the promoter
2 functions in a prokaryote.

1 31. An expression vector as in claim 28 wherein the selectable
2 marker is operably linked to both a prokaryotic and a eukaryotic promoter.

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1  GCTGTCACTT  GGCTCTCTGG  CTGGAGCTTG  AGGACGCAAG  GAGGGTTTGT  CACTGGCAGA
61  CTCGAGACTG  TAGGCACTGC  CATGGCCCTT  GTGCTCAGTA  AGGACTCGGC  GGACATCGAG
121  AGTATCCTGG  CTTTAAATCC  TCGAACACAA  ACTCATGCAA  CTCTGTGTTT  CACTTCGGCC
181  AAGAAATTAG  ACAAGAAACA  TTGGAAAAGA  AATCCTGATA  AGAACTGCTT  TAATTGTGAG
241  AAGCTGGAGA  ATAATTTTGA  TGACATCAAG  CACACGACTC  TTGGTGAGCG  AGGAGCTCTC
301  CGAGAAGCAA  TGAGATGCCT  GAAATGTGCA  GATGCCCCGT  GTCAGAAAGAG  CTGTCCAACT
361  AATCTTGATA  TTAAATCATT  CATCACAAGT  ATTGCAACAA  AGAACTATTA  TGGAGCTGCT
421  AAGATGATAT  TTCTTGACAA  CCCACTTGGT  CTGACTTGTG  GAATGGTATG  TCCAACCTCT
481  GATCTATGTG  TAGGTGGATG  CAATTTATAT  GCCACTGAAG  AGGACCCCAT  TAATATTGGT
541  GGATTGCAGC  AATTGCTAC  TGAGGTATTC  AAAGCAATGA  GTATCCACAA  GATCAGAAAT
601  CCTTCGCTGC  CTCCCCCAGA  AAAAATGTCT  GAAGCCTATT  CTGCAAAAGAT  TGCTCTTTTT
661  GGTGCTGGGC  CTGCAAGTAT  AAGTTGTGCT  TCCTTTTGGG  CTCGATTGGG  GTACTCTGAC
721  ATCACTATAT  TTGAAAACA  AGAATATGTT  GGTGTTTAA  GTACTTCTGA  AATTCCTCAG
781  TTCCGGCTGC  CGTATGATG  AGTGAATTTT  GAGATTGAGC  TAATGAAGGA  CTTGGTGTA
841  AAGATAATT  GCGGTAAAAG  CCTTTCAGTG  AATGAAATGA  CTCTAGCAC  TTTGAAAAGAA
901  AAGGCTACA  AAGCTGCTT  CATTTGCAATA  GGTGTGCCAG  AACCCCAATA  AGATGCCATC
961  TTCCAAGGCC  TGACGCAGGA  CCAGGGGTTT  TATACATCCA  AAGACTTTTT  GCCACTTGTA
1021  GCCAAAGGCA  GTAAAGCAGG  AATGTGCGCC  TGTCACCTCT  CATTGCCATC  GATACGGGGA
1081  GTCGTGATTG  TACTTGGAGC  TGGAGACACT  GCCTTCGACT  GTGCAACATC  TGCTCTACGT
1141  TGTGGAGCTC  GCCGAGTGT  CATCGTCTTC  AGAAAAGGCT  TTGTTAATAT  AAGAGCTGTC
1201  CCTGAGGAGA  TGGAGCTTGC  TAAGGAAGAA  AAGTGTGAAT  TTCTGCCATT  CCTGTCCCA
1261  CGGAAGGTTA  TAGTAAAGG  TGGGAGAATT  GTTGCTATGC  AGTTTGTTCG  GACAGAGCAA
1321  GATGAACTG  GAAAATGGAA  TGAAGATGAA  GATCAGATGG  TCCATCTGAA  AGCCGATGTG
1381  GTCATCAGTG  CCTTTGGTTC  AGTTCTGAGT  GATCCTAAAG  TAAAAGAAGC  CTTGAGCCCT
1441  ATAAATTTA  ACAGATGGGG  TCTCCCAGAA  GTAGATCCAG  AAATAATGCA  AACTAGTGAA
1501  GCATGGGTAT  TTGCAGGTGG  TGATGTCGTT  GGTTCGCTA  ACATAACAGT  GGAATCGGTG
1561  AATGATGGAA  AGCAAGCTTC  TTGGTACATT  CACAAATACG  TACAGTCACA  ATATGGAGCT
1621  TCCGTTTCTG  CCAAGCCTGA  ACTACCCCTC  TTTTACACTC  CTATTGATCT  GGTGGACATT

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FIG. 1A-1.

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1681 AGTGTAGAAA TGGCCGGATT GAAGTTTATA AATCCTTTTG GTCTTGCTAG CGCAACTCCA
1741 GCCACCAGCA CATCAATGAT TCGAAGAGCT TTTGAAGCTG GATGGGGTTT TGCCCTCACC
1801 AAAACTTTCT CTCTTGATAA GGACATTGTG ACAAATGTTT CCCCAGAAAT CATCCGGGA
1861 ACCACCTCTG GCCCCATGTA TGGCCCTGGA CAAAGCTCCT TTCTGAAATAT TGAGCTCATC
1921 AGTGAGAAAA CGGCTGCATA TTGGTGTCAA AGTGTCACTG AACTAAAGGC TGACTTCCCA
1981 GACAACATTG TGATTGCTAG CATTATGTGC AGTTACAATA AAAATGACTG GACGGAACTT
2041 GCCAAGAAGT CTGAGGATTC TGGAGCAGAT GCCCTGGAGT TAAATTTATC ATGTCCACAT
2101 GGCAATGGAG AAAGAGGAAT GGGCCTGGCC TGTGGCAGG ATCCAGAGCT GGTGCGGAAC
2161 ATCTGCCGCT GGGTTAGGCA AGCTGTTTCAG ATTCCTTTT TTGCCAAGCT GACCCCAAAT
2221 GTCACCTGATA TTGTGAGCAT CGCAAGAGCT GCAAAGGAAG GTGGTGCCAA TGGCGTTACA
2281 GCCACCAACA CTGTCTCAGG TCTGATGGGA TTAATACTG ATGGCACACC TTGGCCAGCA
2341 GTGGGGATTG CAAAGCGAAC TACATATGGA GGAGTGTCTG GGACAGCAAT CAGACCTATT
2401 GCTTTGAGAG CTGTGACCTC CATTGCTCGT GCTCTGCCCTC GATTCCCAT TTTGGCTACT
2461 GGTTGGAATTG ACTCTGCTGA AAGTGGTCTT CAGTTTCTCC ATAGTGGTGC TTCCGTCCCTC
2521 CAGGTATGCA GTGCCATTCA GAATCAGGAT TTCACTGTGA TCGAAGACTA CTGCACTGGC
2581 CTCAAAGCCC TGCTTTATCT GAAAAGCATT GAAGAACTAC AAGACTGGGA TGGACAGAGT
2641 CCAGCTACTG TGAGTCACCA GAAAGGGAAC ACCTTATCTG GAACAGCGCA AGAAAATCAT AGCAGAAAAC
2701 AAGAACTGC CAAGTTTGG ACCTTATCTG GAACAGCGCA AGAAAATCAT AGCAGAAAAC
2761 AAGATTAGAC TGAAAGAACA AAATGTAGCT TTTTCACCAC TTAAGAGAAG CTGTTTTATC
2821 CCCAAAAGGC CTATTCCCTAC CATCAAGGAT GTAATAGGAA AAGCACTGCA GTACCTTGA
2881 ACATTTGGTG AATTGAGCAA CGTAGAGCAA GTTGTGGCTA TGATTGATGA AGAAATGTGT
2941 ATCAACTGTG GTAAATGCTA CATGACCTGT AATGATTCTG GCTACCAGGC TATACAGTTT
3001 GATCCAGAAA CCCACCTGCC CACCATAACC GACACTTGTA CAGGCTGTAC TCTGTGTCTC
3061 AGTGTTTGCC CTATTGTGCA CTGCATCAA ATGGTTTCCA GGACAACACC TTATGAACCA
3121 AAGAGAGGCG TACCCTTATC TGTGAATCCG GTGTGTTAAG GTGATTTGTG AAACAGTTGC
3181 TGTGAACCTT CATGTCACCT ACATATGCTG ATCTCTTAAA ATCATGATCC TTGTGTTTCAG
3241 CTCTTTCCAA ATTAAAACAA ATATACATTT TCTAAATAAA AATATGTAAT TTCAAAAATAC
3301 ATTTGTAGT GTAAAAAATG TCTCATGTCA ATGACCATTC AATTAGTGGN CATAAAATAG

FIG. 1A-2.

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3361 AATAATTCTT TTCTGAGGAT AGTAGTTAAA TAACTGTGTG GCAGTTAATT GGATGTTTCCAC
3421 TGCCAGTTGT CTTATGTGAA AAATTAACTT TTTGTGTGGC AATTAGTGTG ACAGTTTCCA
3481 AATTGCCCTA TGCTGTGCTC CATATTGAT TTCTAATTGT AAGTGAAATT AAGCATTTTG
3541 AAACAAAGTA CTCTTTAACA TACAAGAAA TGTATCCAAG GAAACATTTT ATCAATAAAA
3601 ATTACCTTTA ATTTAATGC TGTTTCTAAG AAAATGTAAG TAGCTCCATA AAGTACAAAT
3661 GAAGAAAGTC NAAAATTAT TTGCTATGGC AGGATAAGAA AGCCTAAAAT TGAGTTTGTN
3721 GGACTTTATT AAGTAAATC CCCTTCGCTG AAATTGCTTA TTTTGGTGT TGGATAGAGG
3781 ATAGGGAGAA TATTACTAA CTAATACCA TTCACTACTC ATGCGTGAGA TGGGTGTACA
3841 AACTCATCCT CTTTAAATGG CATTTCTCTT TAAACTATGT TCCTAACCAA ATGAGATGAT
3901 AGGATAGATC CTGGTTACCA CTCTTTTACT GTGCACATAT GGGCCCCGGA ATTC

FIG. 1B.

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1 TCGACCCACG CGTCCGCCGG CCGGAGGCGG AGGACGCGG GAGGGCCCCG CCGTGGGAGA
61 CTCCAAGCTG TCGGCATCGC CATGCCCCCT GTGCTGAGCA AGGACGTGGC GGACATCGAG
121 AGTATCCTGG CTTTAAATCC TCGAACACAG TCTCATGCAG CCCTTCATTC CACTTTGGCC
181 AAGAAATTGG ATAAGAAACA CTGGAAAAGA AATCCCGATA AGAACTGCTT TCATTGCGAG
241 AAGCTGGAGA ATAATTTTGG TGACATCAAG CACACGACTC TTGGTGAGCG AGGAGCTCTC
301 CGAGAAGCAA TGAGATGCCT GAAATGTGCC GATGCTCCCT GTCAGAAAGAG CTGTCCAACT
361 CATCTAGATA TCAAATCATT CATCACAAGT ATCTCAAATA AGAACTATTA TGGAGCTGCT
421 AAGATGATTT TTTCTGACAA CCTCTTGGT CTGACCTGTG GAATGGTATG TCCAACCTCT
481 GATCTTTGTG TAGGAGGATG CAATTTATAT GCAACTGAAG AGGGATCAAT TAATATTGGT
541 GGATTGCAGC AGTTTGCTTC TGAGGTGTTT AAAGCAATGA ATATCCACA AATCAGGAAT
601 CCTTGCTGCG CATCCCAAGA GAAAATGCCT GAAGCTTATT CTGCAAAAGAT TGCTCTTTTG
661 GGTGCTGGGC CTGCAAGTAT AAGCTGTGCT TCCCTTCTGG CTCGATTAGG CTACTCTGAC
721 ATCACTATAT TTGAAAACA AGAATATGTT GGTGGTTTAA GTACTTCTGA AATCCCTCAG
781 TTCCGGCTGC CATATGATGT AGTGAATTT GAGATTGAGC TTATGAAGGA CCTTGGTGTA
841 AAGATAATTT GTGGTAAAG CCTTTCAGAG AATGAATTA CTCTCAACAC TTTAAAGAA
901 GAAGGGTATA AAGCTGCTTT CATTTGGTATA GGTTCGCCAG AACCCAAAC GGATGACATC
961 TTCCAAGGCC TGACACAGGA CCAGGGGTTT TACACATCCA AAGACTTTCT GCCCCTTGA
1021 GCCAAAAGCA GTAAAGCAGG AATGTGTGCC TGTCACCTC CATTGCCATC GATACGGGA
1081 GCCGTGATTG TACTCGGAGC TGGAGACACA GCTTTCGACT GTGCAACATC CGCTTTACGT
1141 TGTGGAGCCC GCCGAGTGT CCTCGTCTTC AGAAAAGGCT TTGTTAATAT AAGAGCTGTC
1201 CCTGAGGAGG TGGAGCTTGC TAAGGAAGAA AAATGTGAAT TTTTGCCCTT CCTGTCCCCA
1261 CGGAAGGTTA TAGTTAAAG TGGAGAAAT GTTGCCGTGC AATTTGTTCC AACAGAACAA
1321 GATGAACTG GAAAATGGAA TGAAGATGAA GATCAGATAG TCCATCTGAA GGCTGATGTG
1381 GTCATCAGTG CCTTTGGCTC AGTGCTGAGG GATCCTAAG GTAGATCCAG AAACATATGA AACCAGTGAA
1441 ATAAAATTA ACAGATGGA TCTCCAGAA GTAGATCCAG AACTATGCA AACCAGTGAA
1501 CCATGGGTGT TTGCAGGTGG TGATATCGTT GGTATGGCTA ACACACGCT GGAATCCGTA
1561 AATGACGGAA AGCAGGCCTC CTGGTACATT CACAAATATA TCCAGGCCCA ATATGGAGCT
1621 TCAGTTTCTG CCAAGCCCCG ACTGCCCCCTG TTTTATACGC CTGTTGACCT GGTGGACATC

FIG. 2A-1.

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1681 AGCGTGGAAA TGGCTGGATT AAAGTTTATA AATCCTTTTG GTCTTGCCAG TGCAGCTCCA
1741 ACTACCAGTT CATCGATGAT TCGAAGAGCT TTTGAAGCTG GATGGGTTT TGCCCTGACC
1801 AAAACTTTCT CTCTTGATAA GGACATAGTG ACAAATGTCT CACCCAGAAAT CGTCCGGGGG
1861 ACTACCTCTG GCCCCATGTA CGGCCCTGGA CAAAGCTCCT TCCTGAATAT TGAGCTCATC
1921 AGTGAAAAAA CAGCTGCATA TTGGTGTCAA AGTGTCAC TG AACTAAAAGC TGACTTTCCA
1981 GACAATATTG TGATCGCCAG CATCATGTGT AGTTACAACA AAAATGACTG GATGGAAC TC
2041 TCCAGAAAGG CTGAGGCCCTC TGGAGCAGAT GCCTTGGAGT TAAATCTGTC ATGTCCACAC
2101 GGCATGGGAG AAAGAGGAAT GGGCCTGGCT TGTGGGCAGG ATCCAGAGCT GGTGCGGAAC
2161 ATCTGTCGCT GGGTTAGGCA AGCTGTTTCAG ATTCCCTTTT TTGCCAAGTT GACCCCAAAC
2221 GTCAC TGATA TAGTAAGCAT CGCCAGAGCG GCCAAGGAAG GTGGCGCAGA TGGTGTACA
2281 GCCACCAACA CGGTCTCAGG TCTCATGGGA TTAAGAGCCG ATGGCACGCC CTGGCCAGCG
2341 GTGGTGCTG GCAAGCGGAC TACATACGGA GGAGTGTCTG GCACGGCCAT CAGACCAATT
2401 GCTTTGAGAG CTGTGACCAC CATTGCTCGT GCTTTGCCCTG GATTCCCAT TTTGGCTACT
2461 GGTGGAATTG ACTCAGCTGA AAGTGGACTT CAGTTTCTCC ACAGTGGTGC TTCGGTCCCTC
2521 CAGGTATGCA GTGCTGTTCA GAATCAGGAT TTCACTGTCA TCCAGACTA TTGCACTGGC
2581 CTCAAAGCCT TGCTTTATCT GAAAAGCATT GAAGAACTAC AAGGCTGGA AGGCGCAGAGT
2641 CCAGGTACCG AGAGTCACCA GAAGGGGAAA CCAGTTCCTC GTATTGCTGA ACTCATGGGA
2701 AAGAACTGC CAAATTTTGG ACCTTATCTG GAGCAACGCA AGAAAATCAT AGCAGAGGAA
2761 AAGATGAGAC TGAAGAACA AAATGCAGCT TTCCACCAC TTGAGAGAAA ACCTTTTATT
2821 CCCAAAAAGC CTATTCTGCTG TATTAAGGAT GTAATTGGAA AAGCACTGCA GTACCTTGGA
2881 ACGTTTGGTG AACTGAGCAA CATAGAGCAA GTTGTGGCTG TGATCGATGA AGAAATGTGT
2941 ATCAACTGTG GCAAATGCTA CATGACCTGT AATGACTCTG GCTACCAGGC TATCCAGTTT
3001 GATCCCGAAA CCCACCTGCC CACCGTTACT GACACTGCA CAGGCTGTAC CCTGTGTCTC
3061 TCCGTCTGCC CTATTATCGA CTGCATCAGA ATGGTTTCCA GGACAACACC TTACGAACCA
3121 AAGAGAGGCT TGCCCTTTGGC TGTGAATCCG GTGTGCTGAG GTGATTCGTG GAACAGTTGC
3181 TGTGAAC TTT GAGGTCACCC CCATATGCTG TCTTTTAAAT TGTGGTTATT ATACTCAGCT
3241 CTTTCTCAAT GAAAAACAAAT ATAATATTTC TAGATAAAAAG TTCTAAATAC ATGTCTAAAT
3301 TTTAAAAAAC ATCTACTGCC AGAGCCCCGTT CAATTAATGG TGATAAAATA GAATCCTCCT

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FIG. 2A-2.

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3361 TTTCTGAGGC TAGTTGTTCA ATAACGTCTG CAGTTAATTG GATGTTCTCC ATCAGTTATC
3421 CATTATGAAA AATATTAACT TTTTGGTGG CAATTTCCAA ATTGCCCTAT GCTGTGCTCT
3481 GTCCTTGATT TCTAATTGTA AGTGAAGTTA AGCATTTTAG AACAAAGTAT AATTTAACTT
3541 TCAAGCAAAT GTTCCCAAGG AACATTTTA TAATTAAAAA TTACAATTTA ATTTTAACAC
3601 TGTTCCTAAG CAAATGTAAT TAGCTCCATA AAGCTCAAAT GAAGTCAAAT AATTATTTAC
3661 TGTGGCAGGA AAAGAAAGCC AATGAGGGTT TGCAAAACTT CTCTAAGGCC CTTTGGCTGA
3721 AATAACTTCT CTTTGGTGCT ACATACTGAA AGTGACTGTT TAATCATCAT TCATGTCACA
3781 CCGTGCTCCC TCGCCCTCAG GCCTGAGATG GGCTCCAGA CTCCACCACT GAATCAGCAT
3841 GACACCTTCT TTAACCTGTG GAGCGACGTT CCTAACAAAG TAAGGTGTGG GGATGAAGCT
3901 CTGGTTAAAG CCACCTCTTT GCTGTGCTCC GATCTGTTCT ATCCGCTTCT GAGAGCAACC
3961 TTCATGATTA CAGCAATTAA TGTTTGCACA GAGCCCAGAT TATACAGCAG TGGGTCATTG
4021 TGCTTCATTA TTCAAGAATG AAGATAAAGA CAAATAGAGG ATTAGTAAAA TATATTAAT
4081 GTGCAATACC ACTTAAATGA CTCCTAATGT TTATATTGAA TTTCCAAAGC GATTAAATAA
4141 AAAAGAGCTA TTTTGTGTTA TTGCCAAACA ATATTTTGTG TATTTCTCTA TTTTCATAAT
4201 GAGCAAAATAG CATCCTATAA ATCTGTTTAT CTCCTCTTTG TAGTGTGTTT TCATATAAAT
4261 CCACAAAGTAG AAAATCTTTT CATCTGTGGC ATATTTCTAT GACAAATGCA AGATCTAGAA
4321 AAATTAAATG TTTGATTATG CCATTTTGGG AATGCATATT TACCACCAAA CCTATGTGAC
4381 TGAATAATGT CAAATAAAAT TTTATGAATC ATTTTAAAAA AAAAAAAAAG AGGGCGGCCG
4441 C

FIG. 2B.

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MAPVLSKDVADIESILALNPRTQSHAALHSTLAKKLDKKHKWRNPDKNCFHCEKLENNFGDIKHTTLGER 70
S T T C S N D
GALREAMRCLKADAPCQKSCPTHLDIKSFITSISNKNYYGAAMIFSDNPLGLTCGMVCPTSDLCVGGC 140
N A
NLYATEGSINIGGLQFASEVFKAMNIPQIRNPCLPSEQKMPAYSAKIALLGAGPASISCASFLARLG 210
P T S S PP S F
YSDITIFEKQEVVGLSTSEIPQFRLPYDVVNFEIELMKDLGVKII CGKSLSENEITLNTLKEEGYKAAF 280
V M S K
NADPH/NADP
IGIGLPEPKTDDIFQFLTQDQGYTSKDELPLVAKSSKAGMCACHSP LPSIRGAVIVLGAGDTAFDCATS 350
NK A G V
ALRCGARRVFLVRKGFVNIRAVPEEVELAKEEKECEFLPFLSPRKVI V KGRIVAVQFVRTEQDET GKWN 420
I M
FAD
EDEDQIVHLKADVVISAFGSVL RDPKVKEALSP IKFN RWDLPEDVDPETMQ TSEPWFAGD I VGMANTTV 490
M S G A V L
ESVNDGKQASWYIHKYIQAQYGASVSAKPELPLFYTPVDLVDISVEMAGLKF INPFGLASAAPTTSSMI 560
V S I T A T
RRAFEAGWGFAITKTFSLDKDIVTNVSPRIVRGTTSGPMYGPQSSFLNI ELISEKTAAYWCQSVTELKA 630
I
URACIL
DFPDNIVIASIMCSYNKNDWMELSRKAESGADALENLSCPHGMGERGMGLACGQDPPELV RNICRWVRQ 700
T AK S D

FIG. 3-1.

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FIG. 3-2.

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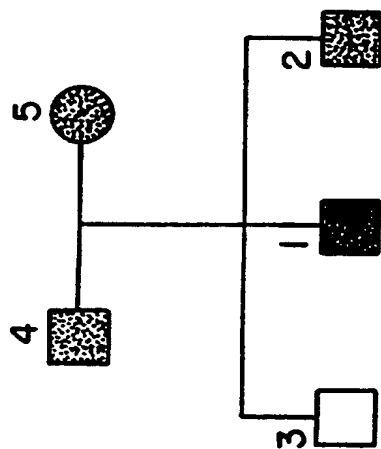


FIG. 4.

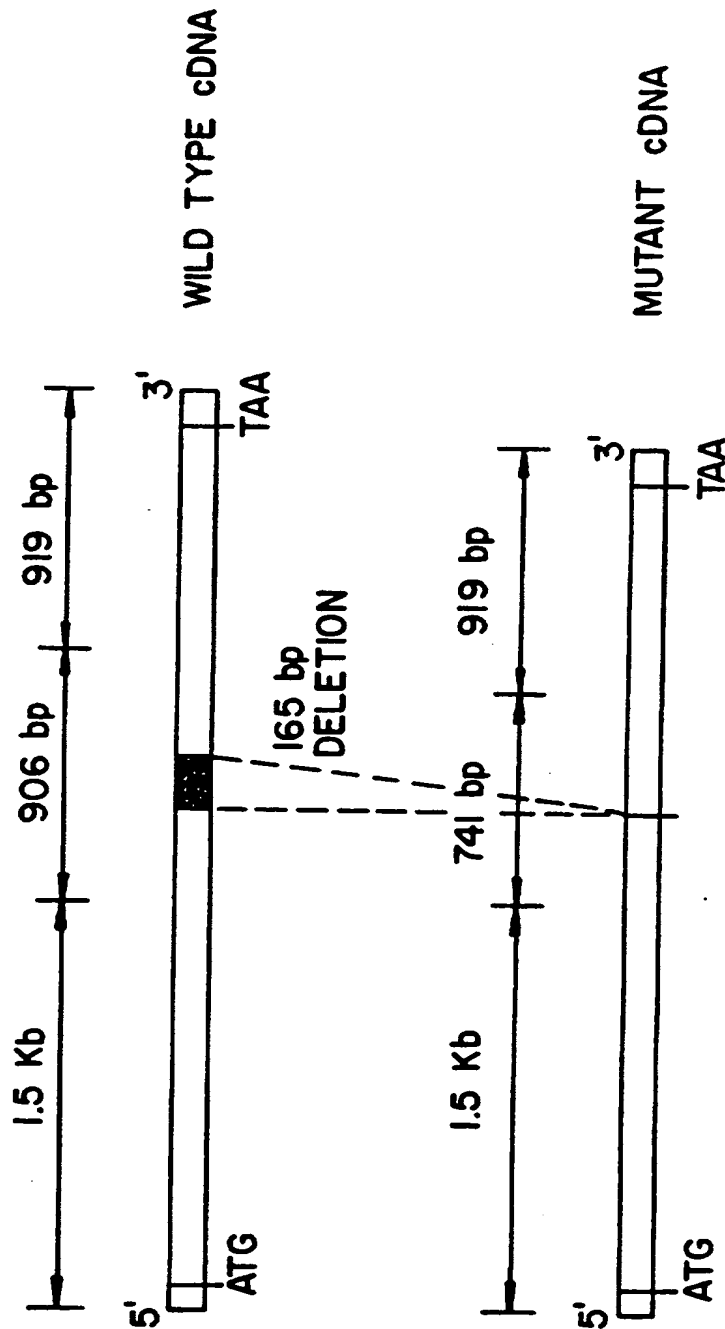
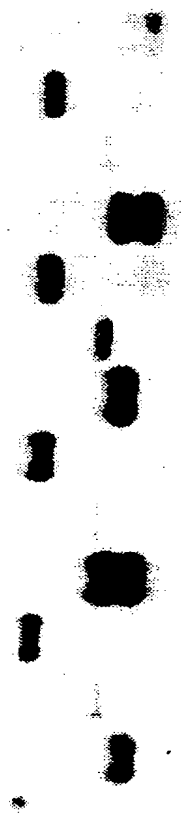


FIG. 6.

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1.5 Kb
906 bp



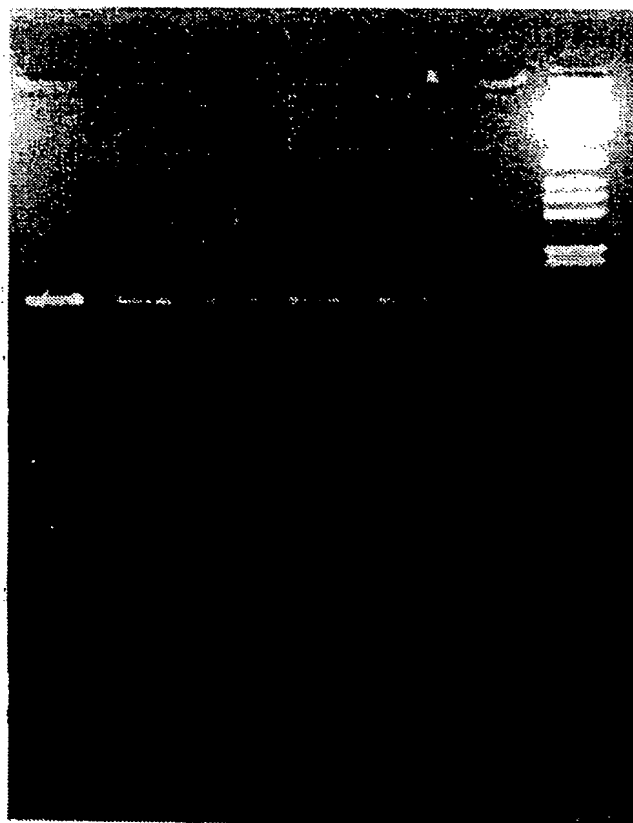
919 bp
741 bp

Subject	1	2	3	4	5
Phenotype	--	+	++	+	+
Genotype	-/-	+/-	+/+	+/-	+/-

FIG. 5.

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2 3 1 4 5 6 7



GENOTYPE

-/-

FIG. 7.

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(21) International Application Number: PCT/US95/12016 (22) International Filing Date: 7 September 1995 (07.09.95) (30) Priority Data: 08/304,309 12 September 1994 (12.09.94) US (71) Applicant: THE GOVERNMENT OF THE UNITED STATES OF AMERICA, represented by THE SECRETARY, DEPARTMENT OF HEALTH AND HUMAN SERVICES [US/US]; Box OTT, Bethesda, MD 20892 (US). (72) Inventors: GONZALEZ, Frank, J.; 5000 Battery Lane #101, Bethesda, MD 20814 (US). FERNANDEZ-SALGUERO, Pedro; 4863 Battery Lane #22, Bethesda, MD 20814 (US). (74) Agents: KRUSE, Norman, J. et al.; Townsend and Townsend and Crew, Stewart Street Tower, One Market, San Francisco, CA 94105-1492 (US).		(81) Designated States: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ, UG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i> (88) Date of publication of the international search report: 17 May 1996 (17.05.96)
(54) Title: CLONING AND EXPRESSION OF cDNA FOR HUMAN DIHYDROPYRIMIDINE DEHYDROGENASE (57) Abstract <p>The invention relates to methods and compositions that are useful for detecting deficiencies in dihydropyrimidine dehydrogenase (DPD) levels in mammals including humans. Cancer patients having a DPD deficiency are at risk of a severe toxic reaction to the commonly used anticancer agent 5-fluorouracil (5-FU). Claimed are DPD genes from human and pig, methods for detecting the level of nucleic acids that encode DPD in a patient, and nucleic acids that are useful as probes for this purpose. Also claimed are methods for expressing DPD in heterologous organisms. Expression vectors that employ a DPD nucleic acid as a selectable marker are also claimed. This selectable marker functions in both prokaryotes and eukaryotes.</p>		

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INTERNATIONAL SEARCH REPORT

International Application No.

PC/US 95/12016

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/53 C12N9/02 C12Q1/68 C12N15/52 C12N15/74
C12N15/79

According to International Patent Classification (IPC) or to both national classification and IPC

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IPC 6 C12N

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Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	J. BIOL. CHEM., vol. 267, no. 24, 1992 pages 17102-17109, LU ET AL 'Purification and characterization of dihydropyrimidine dehydrogenase from human liver' see the whole document ---	1-31
Y	WO,A,92 13077 (BRIGHAM & WOMENS HOSPITAL) 6 August 1992 see page 22, line 20 - page 30, line 3; claims 13,14,17 --- -/--	1-31

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Date of the actual completion of the international search

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Date of mailing of the international search report

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	CANCER RESEARCH , vol. 53, 1993 pages 5433-38, LU ET AL. 'Dihydropyrimidine dehydrogenase activity in human peripheral blood mononuclear cells and liver: population characteristics, newly identified deficient patients, and clinical implication in 5-fluorouracil chemotherapy" ' see the whole document ---	1-31
P,X	J. BIOL. CHEM., 1994, 269, 23192-6, YOKOTA, HIROSHI ET AL 'cDNA cloning and chromosome mapping of human dihydropyrimidine dehydrogenase, an enzyme associated with 5-fluorouracil toxicity and congenital thymine uraciluria' see the whole document ---	1-12
E	WO,A,95 28489 (UAB RESEARCH FOUNDATION) 26 October 1995 see the whole document -----	1-31

INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

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WO-A-9213077	06-08-92	AU-B- 1337092 US-A- 5272078	27-08-92 21-12-93
WO-A-9528489	26-10-95	AU-B- 2289495	10-11-95

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